

Anti-viral Activity of 62 Medicinal Plants, Herbs and Spices Available in Bangladesh: A Mini Review

Adiba Firuj¹, Fahima Aktar¹, Tahmina Akter², Jakir Ahmed Chowdhury³, Abu Asad Chowdhury¹, Shaila Kabir¹, Sultan Mehtap Büyüker⁴ and Md. Shah Amran¹

¹Molecular Pharmacology and Herbal Drug Research Laboratory, Department of Pharmaceutical Chemistry
Faculty of Pharmacy, University of Dhaka, Dhaka-1000, Bangladesh

²Department of Physiology, Dhaka Medical College, Dhaka-1000, Bangladesh

³Department of Pharmaceutical Technology, Faculty of Pharmacy, University of Dhaka
Dhaka-1000, Bangladesh

⁴Department of Pharmacy Services, Üsküdar University, Üsküdar, Istanbul, Turkiye

(Received: February 2, 2023; Accepted: June 14, 2023; Published (web): July 7, 2023)

ABSTRACT: Microorganisms can cause devastating diseases leading to a pandemic, such as COVID-19 which has created a devastating situation throughout the world. The SARS-CoV-2 or severe acute respiratory syndrome coronavirus-2 is the major concern for creating this pandemic situation. Viruses are the major pathogenic microorganisms that have the potential to be detrimental to human beings and animals in many ways. By entering the human body through different routes and commanding different machinery of the body to generate a higher number of their genomic copies and proteins they create pathogenesis. Phytomedicines may act as suitable weapons to halt the reproduction of such viruses and combat diseases caused by these viruses. Bangladesh is the habitat of about 500 medicinal and aromatic plants. Many of these plants have been found to show anti-viral activity. The anti-viral activity of *Terminalia chebula*, a well-known Bangladeshi medicinal plant has been proven to combat Newcastle Disease virus, Herpes Simplex Virus, Adenovirus type 5, Measles virus, Echovirus type 11, Rotavirus, Influenza A virus, Hepatitis B virus and Enterovirus. This review article was aimed to search available medicinal plants as novel anti-viral drugs.

Key words: Medicinal and aromatic plants, Viral diseases, Anti-viral activity of medicinal plant, Newcastle Disease virus, New drug development

INTRODUCTION

Medicinal plants are a precious blessing from nature to humans to help them live healthy and disease-free life. The latest pharmacological research found that medicinal and herbal plants possess anti-viral properties.¹ Thus, by nature we are blessed with several anti-viral herbs that have broad anti-viral spectrum properties.²

Viruses are small biological organisms found practically everywhere. They can infect humans, live stocks, plants, fungi and microbes as well. A virus

can cause many diseases that can be lethal. A virus's influence on different creatures varies, as does its complexity. They contain ribonucleic acid (RNA) or deoxyribonucleic acid (DNA) covered by protein, lipid (fatty components), or glycoprotein (figure 1). They are parasitic because they cannot reproduce without a host.^{3,4}

Viruses have the power to induce a ton of diseases in humans and other living organisms.^{5,6} Among these diseases are smallpox, flu, measles, mumps, chicken pox, hepatitis, sore throat, polio, rabies, ebola virus disease, AIDS (caused by human immunodeficiency virus), respiratory infections, dengue, Epstein-Barr and Guillain-Barre syndrome (caused by cytomegalovirus), etc. Human papillomavirus (HPV) is a type of virus that can cause

Correspondence to: Md. Shah Amran
Email: amranms@du.ac.bd; Mobile: + 8801718-617915
<https://orchid.org/0000-0001-6207-1240>

Dhaka Univ. J. Pharm. Sci. 22(2): 213-232, 2023 (December)
DOI: <https://doi.org/10.3329/dujps.v22i2.67408>

cancer. We will be focusing on the anti-viral characteristics of plants that are cultivated or native to

Bangladesh in this article.

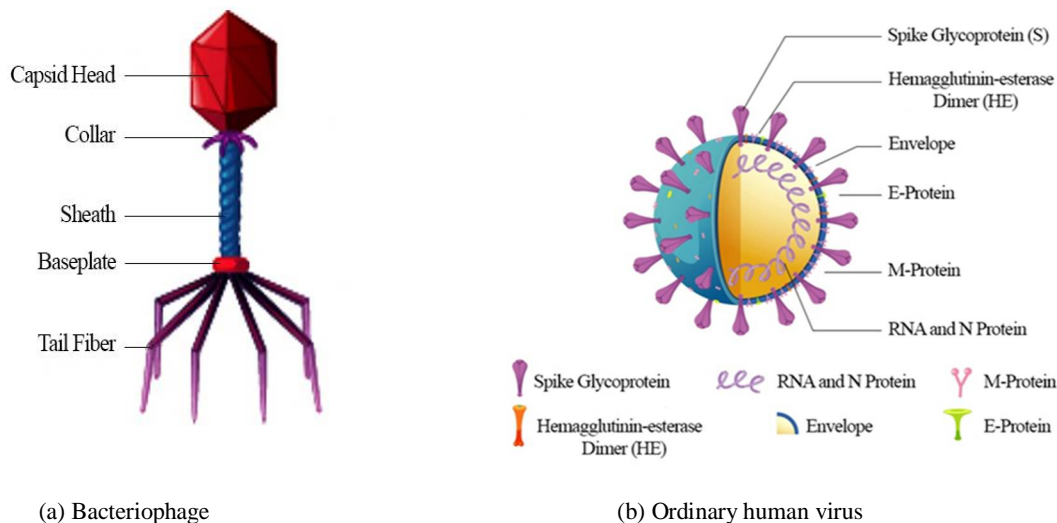


Figure 1. Schematic diagram of the structure of a virus.

MATERIALS AND METHODS

Different electronic sources of databank were searched for the review of the article. Among the sources some are - PubMed, Springer link, MDPI link, ResearchGate, Google and Google Scholar and Science Direct etc.

List of medicinal plants for anti-viral activity.

Plants are an excellent source of very novel chemical compounds that can combat bacteria, viruses, fungi, and other microorganisms. In the context of COVID-19 we searched for medicinal plants that have anti-viral activities. These plants may be good anti-viral sources to combat COVID-19 since they possess anti-viral action against several viral infections. The Ayurvedic Pharmacopeia of Bangladesh has published a detailed description of some indigenous medicinal plants of Bangladesh. The local names of the plants have been written in romaji followed by their botanical name. A list of the searched medicinal plants having anti-viral activity is shown in table 1.

Anti-viral potential of specific plant. Medicinal plants are used for viral infections since ancient times. Extracts collected from leaves, seeds, roots and different parts of medicinal plants have been applied against human viruses. Reviews of several studies are pieces of evidence of the fact that hundreds of medicinal plants possess strong potential against viruses. As the study points out about the potential benefits of antiviral plants, after careful investigations these plants extracts can be used for alleviating different viral infection symptoms including diseases like COVID-19. The anti-viral action of methanolic extracts *Terminalia chebula* was examined in BHK cells against the NDV. The physiological changes generated by NDV in BHK cell lines are the same as those caused by an infection with HIV, which results in the creation of a syncytium by cell-to-cell fusion. These viruses are similar to COVID-19 as the virus induces the formation of syncytia (figure 2)³. Thus *T. chebula* has potential anti-viral activity against the coronavirus.³

Table 1. List of 62 medicinal plants searched for anti-viral activity.

1	Ada (<i>Zingiber officinale</i> Roxb.)	32	Kantakari (<i>Solanum xanthocarpum</i> Schrad. & Wendl.)
2	Akand (<i>Calotropis procera</i>)	33	Karbee (<i>Nerium indicum</i> Mill., Syn. <i>N. odorum</i> Soland.)
3	Am (<i>Mangifera indica</i> Linn.)	34	Katphal (<i>Myrica esculenta</i> Buch. / <i>M. nagi</i> Hook. f.)
4	Amalaki (<i>Phyllanthus embelica</i> / <i>Embelica officinalis</i>)	35	Khayer (<i>Acacia catechu</i> Linn. f.)
5	Aparajita (<i>Clitoria ternatea</i>)	36	Khude dhekishak (<i>Ceratopteris thalictroides</i>)
6	Arjuna (<i>Terminalia arjuna</i>)	37	Kurchi (<i>Holarrhena antidysenterica</i> Roth.)
7	Ashoka (<i>Saraca asoka</i> Rose/ <i>Saraca Indica</i> Linn)	38	Kusum fool (<i>Carthamus tinctorius</i>)
8	Ashwagandha (<i>Withania somnifera</i> Dunal)	39	Kutki [Birhalnokha](<i>Capparis spinosa</i>)
9	Bahera (<i>Terminalia bellerica</i> Roxb.)	40	Kutumkanta (<i>Caesalpinia crista</i>)
10	Banar-mul (<i>Vetiveria zizanioides</i> Linn.)	41	Lajjabati (<i>Mimosa pudica</i> Linn.)
11	Barun (<i>Crataeva nurvala</i> Buch-Ham.)	42	Methi (<i>Trigonella foenum-graecum</i> Linn.)
12	Basak (<i>Adhatoda vasica</i> Nees.)	43	Morog fool (<i>Celosia cristata</i>)
13	Bathua (<i>Chenopodium album</i>)	44	Mutha (<i>Cyperus rotundus</i>)
14	Bel (<i>Aegle marmelos</i> Corr.)	45	Neem (<i>Azadirachta indica</i> A. Juss.)
15	Bherenda (<i>Ricinus communis</i> Linn.)	46	Nishinda (<i>Vitex negundo</i> Linn.)
16	Bhui-kumra (<i>Pueraria tuberosa</i> DC.)	47	Patte-madar (<i>Erythrina indica</i> Lam. / <i>E. variegata</i> L.)
17	Bhumi amalaki (<i>Phyllanthus niruri</i> Hook. / <i>P. fraternus</i> webst.)	48	Pipul (<i>Piper longum</i> Linn.)
18	Brahmi (<i>Bacopa monnieri</i> Linn.)	49	Punarnava (<i>Boerhavia diffusa</i>)
19	Calendula fool (<i>Calendula officinalis</i>)	50	Randhuni, Ajamoda (<i>Apium leptophyllum</i> Pers. / <i>A. graveolens</i> Linn.)
20	Dhutra (<i>Datura metel</i> Linn. / <i>D. fastuosa</i>)	51	Rasna (<i>Pluchea lanceolata</i> Oliver & Hiern.)
21	Drun-puspi (<i>Leucas cephalotes</i> Spreng. / <i>L. aspera</i> Spreng.)	52	Rasun (<i>Allium sativum</i> Linn.)
22	Durba (<i>Cynodon dactylon</i> Linn.)	53	Sajna (<i>Moringa oliefera</i> Lam. Syn.)
23	Gamar (<i>Gmelina arborea</i> Roxb.)	54	Sal-parni (<i>Desmodium gangeticum</i> DC.)
24	Gulancha (<i>Tinospora cordifolia</i> Willd.)	55	Sarisa (<i>Brassica campestris</i> Linn. / <i>B. nigra</i> Linn.)
25	Halud (<i>Curcuma longa</i> Linn.)	56	Shimul (<i>Bombax ceiba</i> Linn. / <i>Salmalia malabarrica</i> Sehoff & Endl.)
26	Haritaki (<i>Terminalia chebula</i> Retz.)	57	Sondal (<i>Cassia fistula</i> Linn.)
27	Hurhuriya/Ajagandha (<i>Cleome gynandra</i> Linn.)	58	Supari (<i>Areca catechu</i> Linn.)
28	Jongli fool (<i>Astragalus hamosus</i>)	59	Tajpata (<i>Cinnamomum tamala</i> Buch. Ham.)
29	Kalabati (<i>Canna indica</i>)	60	Tulsi (<i>Ocimum sanctum</i> Linn.)
30	Kalajira (<i>Nigella sativa</i> Linn.)	61	Tut begun (<i>Solanum nigrum</i> Linn)
31	Kanchana (<i>Bauhinia variegata</i> Blume.)	62	Ulkushi (<i>Mucuna prurita</i> Hook. / <i>M. pruriens</i> Baker.)

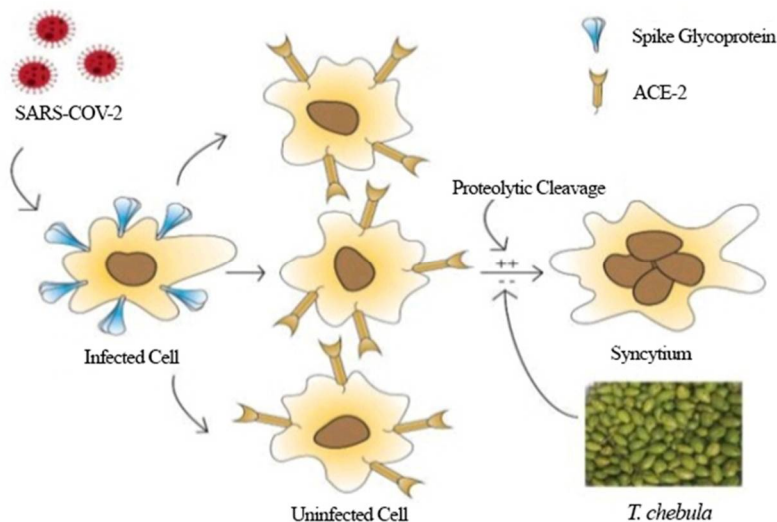


Figure 2. Schematic representation of inhibition of formation of syncytium by *T. chebula*.

Table 2. Medicinal plants and their bio-active compounds for potential anti-viral effects.

Plant species	Bengali name	Family	Part used	Pharmacological action	Bioactive compounds	Effective against
<i>Zingiber officinale</i> Roxb. ⁷⁻¹²	Ada	Zingiberaceae	Rhizome	Nausea, motion sickness, gastric ulcers, diabetes, xerostomia, sore throat, rheumatoid arthritis, migraine pain, and minor respiratory problems	Zingiberol, zingiberene, gingerols, shogaols, 1-dehydro-(10)-gingerdione, terpenoids, flavonoids	Human respiratory syncytial virus (HRSV), feline calicivirus, influenza type A (H ₁ N ₁), hepatitis C virus (HCV), herpes simplex virus type 1 & 2 (HSV-1,2)
<i>Calotropis procera</i> ^{13,14}	Akand	Apocynaceae	Leaves, roots, flowers	Anti-diarrhoeal, anti-stomatitic, used to treat constipation and skin diseases	Cardenolides, terpenes, flavonoids etc.	Foot and mouth disease virus (FMDV)
<i>Mangifera indica</i> Linn. ¹⁵⁻¹⁷	Am	Anacardiaceae	Leaves, root extracts	Antioxidant, anti-viral, anti-diabetic, gastroprotective, cardioprotective, anti-microbial, anti-allergic, antipyretic effects	Amino acids, carbohydrates, fatty acids, vitamins, saponins such as tetrahydroxy pyrrolidone saponin or mangiferin etc.	Influenza virus (H ₉ N ₁), herpes simplex type 1 and type 2 (HSV-1,2)
<i>Phyllanthus Embelica</i> ¹⁸⁻²¹	Amalaki	Phyllanthaceae	Fruits, roots	Antimicrobial, antioxidant, anti-inflammatory, analgesic and antipyretic, adaptogenic, hepatoprotective, antitumor and antiulcerogenic activities	4'-hydroxy-phyllaemblicin B, nor-sesquiterpenoid glycosides, phyllaemblicin A, phyllaemblicin B and phyllaemblicin C, phyllaemblicin E and F, methyl ester of phyllaemblic acid, 1,2,4,6-tetra-O-galloyl-D-glucose (1246TGG)	Coxsackievirus B ₃ , human immunodeficiency virus (HIV), HSV (1 & 2)
<i>Clitoria Ternatea</i> ^{22,23}	Aparajita	Fabaceae	Flowers, leaves, seeds, stems, roots	Analgesic, insecticidal, antioxidant, anticancer activities	Flavonoids, alkaloids, anthraquinone, anthocyanins, volatile oils, steroids, stigmast-4-ene-3,6-dione	Bacteria, yeast, fungi
<i>Terminalia Arjuna</i> ²⁴⁻²⁶	Arjuna	Combretaceae	Bark extracts	Anti-hypertensive, hypolipidemic, anti-thrombotic, anti-coagulant, antiviral, anti-bacterial, and anti-fungal effects	arjungenin, arjunin, arjunic acid, arjunolic acid, terminic acid, casuarinin, casurin, punicalagin, castalagin, asarjunone, arjunolone, luteolin, bicalein, gallic acid	Herpes simplex virus type 2 (HSV-2)
<i>Saraca asoka</i> Rosc/ <i>Saraca Indica</i> Linn. ^{27,28}	Ashoka	Fabaceae	Leaves	Spasmogenic, anti-tumor, anti-cancer, oxytocic, anti-bacterial, anti-implantation, and menorrhagic activities	Tannins, flavonoids, carbohydrates, proteins, steroids	<i>Bacillus subtilis</i> , <i>Pseudomonas aeruginosa</i> , <i>Candida albicans</i> , <i>Aspergillus niger</i>
<i>Withania somnifera</i> Dunal ²⁹⁻³¹	Ashwagandha	Solanaceae	Leaves, fruits, bark	Anti-viral, anti-microbial, anti-inflammatory, neuroprotective, cardioprotective, anti-tumor, and anti-diabetic effects	Withaferin A	Influenza virus (H ₁ N ₁), HSV-1, infectious bursal virus disorder
<i>Terminalia bellerica</i> Roxb. ³²⁻³⁴	Bahera	Combretaceae	Fruits	Anti-diabetic, anti-microbial, anti-bacterial, anti-fungal, anti-cancer, hepatoprotective, antipyretic, anti-diarrheal effects	Tannins, glycosides, flavonoids, phenolic compounds, amino acids, saponins	Newcastle disease virus (NDV), human papillomavirus (HPV), human immunodeficiency virus (HIV)

<i>Vetiveria zizanioides</i> Linn. ³⁵⁻³⁷	Banar-mul	Poaceae	Roots	Anti-bacterial, insecticidal, herbicidal, and antioxidant properties	Khushimol, α -vetiverone, β -vetiverone, ethyl 4-(4-methylphenyl)-4-pentenoate	Dengue virus
<i>Crataeva nurvala</i> Buch-Ham. ^{38,39}	Barun	Capparaceae	Leaves	Used to treat inflammatory diseases, paralysis, urogenital problems, nephrotic disorders, breast cancer, thyroid problems	Lupeol, cadabicine, lupenone, mannitol, phragmalin triacetate, succinic acid, betulinic acid, lactic acid, stigmasterol, kaemferol-3-O-D-glucoside, quercetin-3-O-D-glucoside	Bacteria, fungi
<i>Adhatoda vasica</i> Nees. / <i>Justicia adhatoda</i> L. ⁴⁰⁻⁴³	Basak	Acanthaceae	Leaves, roots, bark	Anti-inflammatory, antioxidant, anti-fungal, anti-viral, antitussive, anti-ulcer, thrombolytic, hepatoprotective, and cardioprotective properties	vasicine, l-vasicinone, deoxyvasicine, maiontone, vasicinolone and vasicinol	Influenza virus, human immunodeficiency virus (HIV)
<i>Chenopodium album</i> ^{44,45}	Bathua	Amaranthaceae	Leaves	Used to treat tumors, cancer, headaches, bacterial and fungal infections, amoebiasis, inflammation, also used for contraception, diarrhea, hepatic disorders, diabetes, and immune disorders	alkaloids, flavonoids, anthocyanidine, saponins, glycosides, tannins, carbohydrates	Tobacco mosaic virus (TMV), sunn hemp rosette virus (SRV)
<i>Aegle marmelos</i> Corr. ⁴⁶⁻⁵⁰	Bel	Rutaceae	Leaves, fruits, bark	Antidiarrhoeal, antimicrobial, antiviral, radioprotective, anticancer, and chemopreventive effects, antipyretic, ulcer healing, antigenotoxic, diuretic, anti-fertility and anti-inflammatory properties	Marmelide, Steroids, terpenoids, flavonoids, phenolic compounds, lignin, fat and oil, inulin, proteins, carbohydrates	Human coxsackievirus B ₃
<i>Ricinus communis</i> Linn. ⁵¹⁻⁵³	Bherenda	Euphorbiaceae	Leaves, fruits	Antioxidant, antihistaminic, Antinociceptive, antiasthmatic, antiulcer, anti-diabetic, hepatoprotective, Antifertility, anti-inflammatory, antimicrobial, central nervous system stimulant, lipolytic, wound healing, insecticidal and larvicidal properties	Steroids, saponins, alkaloids, flavonoids, glycosides, ricinine, N-demethylricinine, gallic acid, quercetin, gentisic acid, rutin, epicatechin, ellagic acid	Coxsackievirus B ₃ , HSV-1, vesicular stomatitis virus (VSV)
<i>Pueraria tuberosa</i> DC. ^{54,55}	Bhui-kumra	Fabaceae	Leaves, tuber	Antioxidant, anti-inflammatory, wound healing effects, anticancer, anti-diabetic, contraceptive, anti-convulsant, anxiolytic, anti-ulcer, hepatoprotective, cardioprotective, lipid-lowering, neuroprotective, nephroprotective properties	Biochanin A and B, puerarin, genistein, daidzein, quercetin, irisolidone, isoorientin, mangiferin	<i>Escherichia coli</i> , <i>Staphylococcus aureus</i> , <i>Salmonella paratyphi</i> , <i>Bacillus cereus</i> , <i>Candida albicans</i> , <i>Alternaria solani</i> , <i>Aspergillus fumigates</i>
<i>Phyllanthus niruri</i> Hook. ⁵⁶⁻⁵⁹	Bhumi amalaki	Phyllanthaceae	Leaves	Used for jaundice, gonorrhoea, frequent menstruation, and diabetes, topically used as a poultice	4-O-caffeoylquinic acid, quercetin-3-O-rhamnoside	Hepatitis C Virus (HCV), Hepatitis B virus (HBV), Woodchuck

				for skin ulcers, sores, swelling, and itchiness		Hepatitis virus (WHV), Human immunodeficiency virus type-1 (HIV-1)
<i>Bacopa monnieri</i> Linn. ⁶⁰	Brahmi	Scrophulariaceae	Leaves, flowers	Used for memory enhancement, improvement of concentration and learning, relieves anxiety and epilepsy, cardiostimulant, gastroprotective, hepatoprotective effects	alkaloids, brahmine, herpestine, saponins, d-mannitol, hersaponin, monnierin, bacosides, bacopasaponins	Bacteria, fungi
<i>Calendula officinalis</i> ^{61,62}	Calendula fool	Asteraceae	Leaves, flowers	Antioxidant, anti-microbial, anti-proliferative, anti-inflammatory, anti-ulcer, glucose controlling, lipid-lowering, and wound healing properties	Phenolic compounds, flavonoids, carotenoids, fatty acids, saponins, amino acids, steroids, carbohydrates	Human immunodeficiency virus type-1 (HIV-1)
<i>Datura metel</i> Linn. / <i>D. fastuosa</i> ⁶³⁻⁶⁶	Dhutra	Solanaceae	Fruits, seeds, flowers	Treats asthma symptoms, epilepsy, hysteria, insanity, heart and skin diseases, mental disorders	Atropine, hyoscyamine, and scopolamine alkaloids, tannin, phenol, lignins, lipids, proteins, saponin, glycosides, sterols	Rabies virus (RV)
<i>Leucas aspera</i> Spreng. ^{67,68}	Drun-puspi	Lamiaceae	Leaves, stem, roots, flowers	Antipyretic, insecticidal, anti-fungal, antioxidant, anti-microbial, anti-nociceptive, cytotoxic effects	Alkaloids, steroids, flavonoids, terpenoids, saponins, carbohydrates, proteins, amino acids	Mouse coronavirus (MCV), herpes simplex virus (HSV)
<i>Cynodon dactylon</i> Linn. ⁶⁹⁻⁷³	Durba	Poaceae	Whole plant	Cardiovascular, antidiabetic, gastrointestinal, antioxidant, immunological, antiallergic, anti-inflammatory, antipyretic, analgesic, anticancer, dermatological, diuretic, protective, antimicrobial, antiparasitic, insecticidal, and repellent properties	Flavonoids, alkaloids, glycosides, terpenoids, triterpenoids, steroids, saponins, tannins, resins, phytosterols, reducing sugars, carbohydrates, proteins, volatile oils, and fixed oils	White spot syndrome virus (WSSV), porcine reproductive and respiratory syndrome virus (PRRSV), chikungunya virus
<i>Gmelina arborea</i> Linn. ⁷⁴⁻⁷⁶	Gamar	Verbenaceae	Root, bark	Fever, dyspepsia, hallucinations, hyperdipsia, hemorrhoids, gastralgia, burning sensation	Apigenin	<i>Staphylococcus aureus</i> , <i>Escherichia coli</i> , <i>Salmonella typhi</i>
<i>Tinospora cordifolia</i> Willd. ^{77,78}	Gulanha	Menispermaceae	Leaves, fruits, seeds	Antipyretic, anti-diarrhoeal, anti-leprotic, anti-diabetic, nephroprotective and healing effects for skin diseases	Alkaloids, terpenoids, lignans, steroids	Hepatitis A virus (HAV)
<i>Curcuma longa</i> Linn. ⁷⁹⁻⁸⁵	Halud	Zingiberaceae	Rhizome	Anti-inflammatory, antioxidant, antimutagenic, antidiabetic, antibacterial, hepatoprotective, expectorant, anticancer properties	Curcumin, dimethoxy curcumin, bisdemethoxy curcumin, β -turmerone, terpinolene, α -phellandrene, curcumadiol	Hepatitis B virus (HBV), Newcastle disease virus (NDV), dengue virus type 2, influenza virus (H ₁ N ₁ and H ₉ N ₂)

<i>Terminalia chebula</i> Retz. ⁸⁶⁻⁹⁰	Haritaki	Combretaceae	Fruits	Increase appetite, digestive aid, liver stimulant, stomachic, gastrointestinal prokinetic agent, and mild laxative	Chebularic acid, chebulinic acid, pyrogallol, gallic acid, ellagic acid, corilagin, chebulanin, neochebulinic acid, casuarinin	Newcastle disease virus (NDV), hepatitis B virus (HBV), herpes simplex virus (HSV-1,2), influenza A virus (IAV), adenovirus type 5, HIV
<i>Cleome gynandra</i> Linn. ⁹¹⁻⁹⁴	Hurhuriya	Cleomaceae	Leaves, shoots	Anti-inflammatory, anti-cancer, antioxidant, anti-diabetic, and immune modulating properties	Alkaloids, flavonoids, phenols, proteins, amino acids, carbohydrates, steroids, saponins and tannins	<i>Staphylococcus aureus</i> , <i>Candida albicans</i>
<i>Astragalus hamosus</i> Linn. ⁹⁵	Jongli fool	Papilionaceae	Flowers, seed pods	Used for headaches, gastritis, vertigo, stroke, dementia	Flavonoids, fatty acids, steroids, alkene and terpenoids	Hepatitis B virus (HBV)
<i>Canna indica</i> Linn. ^{96,97}	Kalaboti	Cannaceae	Roots, flowers, rhizomes	Anti-bacterial, anti-viral, anthelmintic, anti-inflammatory, analgesic, immunomodulatory, antioxidant, cytotoxic, hemostatic, hepatoprotective, anti-diarrheal properties	Alkaloids, flavonoids, terpenoids, carbohydrates, proteins, cardiac glycosides, steroids, tannins, saponins, phlobatinins	Human immunodeficiency virus type-1 (HIV-1)
<i>Nigella sativa</i> ⁹⁸⁻¹⁰⁴	Kalajira	Ranunculaceae	Seeds	Antioxidant, antidiabetic, immunomodulator, anti-inflammatory, anticancerous activities	Alkaloids, terpenes, phenols, fixed oils, carbohydrates	Hepatitis C virus (HCV), murine cytomegalovirus (MCMV), influenza virus (H ₉ N ₂ & H ₅ N ₁), severe acute respiratory syndrome coronavirus-2 (SARS-CoV-2)
<i>Bauhinia variegata</i> Blume. ^{105,106}	Kanchana	Caesalpiniaceae	Flowers, leaves, bark	Anti-cancer, anti-bacterial, anti-inflammatory, hepatoprotective, hemagglutination properties	Phenolic compounds, terpenoids, tannins, flavonoids	Rotavirus
<i>Solanum xanthocarpum</i> Schrad. & Wendl. ^{107,108}	Kantakari	Solanaceae	roots, stems, leaves, fruits	anti-asthmatic, hypoglycemic, hepatoprotective, anti-bacterial, insecticidal effects	Solasodine, diosgenin	<i>Staphylococcus aureus</i> , <i>Escherichia coli</i> , <i>Pseudomonas aeruginosa</i> , <i>Aspergillus niger</i>
<i>Nerium indicum</i> Mill. ¹⁰⁹⁻¹¹¹	Karabee	Apocynaceae	Flowers, leaves, seeds, bark	Cardioprotective, anti-asthmatic, antioxidant, antiulcer, wound healing, anticancer, antiepileptic properties	Oleandrin, phenols, flavonoids, glycosides	Herpes simplex virus (HSV), HIV
<i>Myrica esculenta</i> Buch. ¹¹²⁻¹¹⁶	Katphal	Myraceae	Bark extracts	Analgesic, anti-allergic, anxiolytic, anti-microbial, anti-diabetic, anti-hypertensive, anti-ulcer, anti-inflammatory properties	Phenols, glycosides, alkaloids, triterpenoids, volatile oils, Dodecanol, phytol, furfurals, 4-H-pyran-4-one	<i>Staphylococcus aureus</i> , <i>Escherichia coli</i> , <i>Streptococcus pyogenes</i>
<i>Acacia catechu</i> Linn.f. ^{117-119, 21}	Khayer	Fabaceae	Stem bark,	Antibacterial, antifungal, anti-diarrhoeal,	Tannins, catechin, phlobatannins,	Newcastle disease virus

			leaves	antioxidant, immunomodulator, antipyretic effects	catechutannic acid, quercetin, quercitrin, fisetin, gums, resins	(NDV), herpes simplex virus type-2 (HSV-2), HIV-1
<i>Ceratopteris thalictroides</i> ¹²⁰⁻¹²³	Khude dhekishak	Pteridaceae	Whole plant	Used as poultice for skin disorders, wound healing properties	Cyanovirin-N, alkaloids, steroids, coumarins, tannins, saponins, flavonoids, quinones, protein, carbohydrate, glycosides, catechin, terpenoids	HIV-1, Ebola virus, influenza viruses
<i>Holarrhena antidysenterica</i> Roth. ¹²⁴⁻¹²⁷	Kurchi	Apocynaceae	Stem bark, seeds	Anti-bacterial, analgesic, free radical scavenging effects, anti-diarrheal, CNS stimulant properties, anti-inflammatory, anti-amoebic, anti-hemorrhoidal, anti-malarial, anti-diabetic, anti-mutagenic effects	Coumarins, flavonoids, steroidal alkaloids, ergosterol, phenolic compounds, tannins, saponins, triterpenoids, resins	Bacteria, fungi
<i>Carthamus tinctorius</i> L. ^{128,129}	Kusum fool	Asteraceae	Flowers	Cardiovascular, blood stasis, cerebrovascular illnesses, anti-bacterial, anti-microbial, and anti-inflammatory properties	Carthamidin, isocarthamidin, safflor yellow A, hydroxysafflor yellow A, luteolin, safflamin C etc.	Kaposi's sarcoma-associated herpes virus (KSHV)
<i>Capparis spinosa</i> ^{130,131}	Kutki	Capparaceae	Fruits, flowers, leaves, bark	Anti-bacterial, antiviral, antifungal, antioxidant, antihypertensive, antidiabetic, lipid-lowering, anti-allergic, hepatoprotective effects	Flavonoids, alkaloids, steroids, terpenoids, tocopherols	Herpes simplex virus type-2 (HSV-2)
<i>Caesalpinia crista</i> ^{132,133}	Kutum-kanta	Caesalpinaceae	Seed	Anti-microbial, antioxidant, anti-cancer, hepatoprotective, antidiabetic, anti-convulsant, insecticidal properties	Flavonoids, alkaloids, tannins, phytosterols, triterpenoids, coumarins	Paramyxovirus, orthomyxovirus
<i>Mimosa pudica</i> Linn. ^{134,135}	Lajjabati	Mimosaceae	Whole plant	Used to treat gastrointestinal issues and sleeplessness, cancer, numerous urogenital infections	Terpenoids, phenolics, flavonoids, glycosides, alkaloids, quinines, tannins, saponins, coumarin	Mumps virus
<i>Trigonella foenum-graecum</i> Linn. ¹³⁶⁻¹³⁸	Methi	Fabaceae	Seeds, leaves	Anti-microbial, hypotensive, antioxidant, anti-inflammatory, and antitumor activity	Sapogenins, isoleucine, galactomannans, flavonoids, phenolic compounds	Bacteria
<i>Celosia Cristata</i> ¹³⁹⁻¹⁴¹	Morog fool	Amaranthaceae	Leaves	Hepatoprotective, lipid-lowering, antimicrobial, anthelmintic, antioxidant, antimelanocyte, anti-aging effects	phenolic compounds, tannins, flavonoids, sterols	Bovine viral diarrhea (BVDV), Hepatitis B virus (HBV)
<i>Cyperus rotundus</i> ¹⁴²⁻¹⁴⁶	Mutha	Cyperaceae	Whole plant	Used for diarrhea, diabetes, inflammation, malaria, digestive and bowel diseases, gastrointestinal issues	Sesquiterpenoids, kaempferol, luteolin, quercetin	Hepatitis B virus (HBV), herpes simplex virus type 1 (HSV-1), Cocksackie virus
<i>Azadirachta indica</i> A. juss. ¹⁴⁷⁻¹⁵¹	Neem	Maliaceae	Leaves, fruits, seeds, stem bark	Treats eczema and acne, wound healing, antidiabetic, anti-inflammatory and antiheperglycemic properties	Azadirachtin A-G, alkaloids, flavonoids, triterpenoids, phenolic compounds, carotenoids, steroids, ketones	Cocksackie B-4 virus, HSV-1, polio virus type-1 (PV-1)

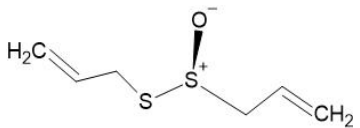
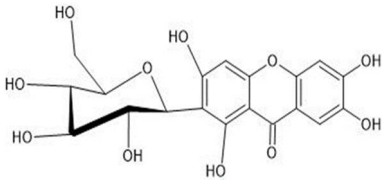
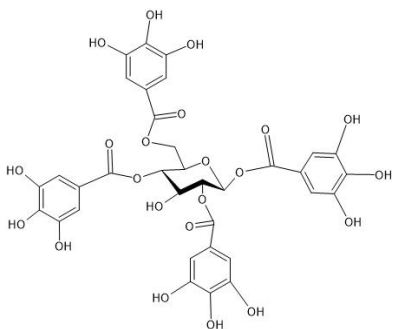
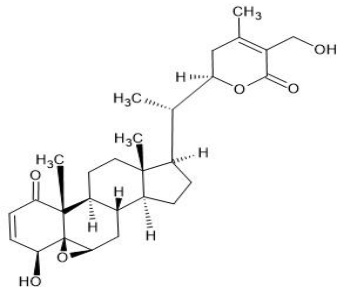
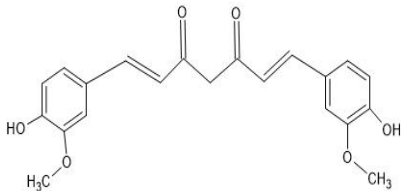
<i>Vitex negundo</i> Linn. ¹⁵²⁻¹⁵⁴	Nishinda	Lamiaceae	Leaves, bark, roots	Antioxidant, anti-inflammatory, analgesic, stimulatory properties, anti-bacterial and anti-tumor properties, anti-arthritic properties, anxiolytic properties, larvicidal, nephroprotective, anti-HIV properties, anti-snake venom properties	Flavonoids, essential oils, flavonoid glycosides, terpenes, lignans, stilbene derivatives	HIV-1, chikungunya virus
<i>Erythrina variegata</i> L. ¹⁵⁵⁻¹⁵⁷	Patte-madar	Fabaceae	Leaves	Anti-asthmatic, anti-epileptic, antiseptic, astringent, anti-inflammatory, analgesic, anthelmintic, smooth muscle relaxant, central nervous system depressive properties	Polyphenolic compounds, flavonoids, triterpenes, gallic acid, caffeic acid	<i>Pseudomonas aeruginosa</i> , <i>Staphylococcus aureus</i> , <i>Escherichia coli</i> , <i>Bacillus subtilis</i> , <i>Bacillus cereus</i>
<i>Piper longum</i> L. ¹⁵⁸⁻¹⁶¹	Pipul	Piperaceae	Roots, seeds, fruits, whole herb	Insecticidal, antifungal, antiamebic, antimicrobial, antiviral, antimicrobial, antioxidant, anticancer, antiasthmatic, hypocholesterolemic, and analgesic activities	Piperine, alkaloids, volatile oils, amides, lignans, esters	Hepatitis B virus (HBV)
<i>Boerhavia diffusa</i> ¹⁶²⁻¹⁶⁵	Punarnava	Nyctaginaceae	Roots, leaves	Analgesic, anti-inflammatory, diuretic, hepatoprotective, immunomodulator, nephroprotective, antiulcer, antihistaminic properties	Punarnavine, boeravinone A-F, punarnavoside, dimethylflavone, boerhavin, boerhavic acid, caffeoyltartaric acid, quercetin, kaempferol	Potato X virus, yellow mosaic virus, bacteriophages
<i>Apium leptophyllum</i> Pers. ¹⁶⁶⁻¹⁶⁸	Randhuni	Apiaceae	Stem, leaves	Asthma, flatulence, dyspepsia, diarrhoea, laryngitis, rheumatoid arthritis, bronchitis	Thymohydroquinone dimethyl ether, thymol methyl ether, isothymol methyl ether, cuminaldehyde, p-cymene, γ -terpinene	<i>Staphylococcus aureus</i> , <i>Escherichia coli</i> , <i>Pseudomonas aeruginosa</i> , <i>Bacillus cereus</i> , and <i>Bacillus subtilis</i> , fungi
<i>Pluchea lanceolata</i> Oliver & Hiern. ¹⁶⁹⁻¹⁷²	Rasna	Asteraceae	Leaves, rhizome	Anti-tumor, wound healing effects, anti-parasitic, anti-malarial, anti-microbial, anti-hyperglycemic, anti-inflammatory effects	Alkaloids, tannins, flavonoids, steroids, phenols, glycosides, triterpenes, proteins, carbohydrates	<i>Vibrio cholerae</i>
<i>Allium sativum</i> Linn. ¹⁷³⁻¹⁷⁶	Rasun	Amaryllidaceae	Bulb	common cold, influenza, hypertension, diabetes	Allitridin, alliin, allicin, diallyl sulfide, diallyl disulfide, diallyl trisulfide, ajoene, and S-allylcysteine	Human cytomegalovirus (HCMV) influenza B virus (IBV), HSV types 1 and 2, parainfluenza virus type 3, vaccinia virus, and human rhinovirus type 2
<i>Moringa oleifera</i> Lam. ¹⁷⁷⁻¹⁸⁴	Sajna	Moringaceae	leaves, fruits, seeds	Used for diabetes, skin infections, anemia, malaria, tuberculosis, headaches, and diseases that are transmitted through sexual contact	Alkaloids, saponins, tannins, steroids, phenolic compounds, glucosinolates, flavonoids, terpenes	Herpes simplex type 1 & type 2 (HSV-1,2), Newcastle disease virus (NDV), human

						immunodeficiency virus (HIV-1), foot and mouth disease virus (FMDV)
<i>Desmodium gangeticum</i> DC. ¹⁸⁵⁻¹⁸⁸	Sal-parni	Fabaceae	Flowers, leaves	Antioxidant, anti-inflammatory, anti-leishmanic, cardioprotective, antiulcer, nephron-protective, wound healing activities	Flavonoids, alkaloids, steroids, terpenoids, pterocarpan, coumarins, volatile oils, phenylpropanoids	Peste des Petits Ruminants (PPR) virus
<i>Brassica nigra</i> Linn. ¹⁸⁹⁻¹⁹²	Sarisa	Brassicaceae	Seeds, flowers, leaves	Cytotoxic, anti-inflammatory, anti-diabetic, mutagenic, hepatoprotective, nephroprotective, anti-obesity, antioxidant, immunomodulator, cardiovascular, hypolipidemic effects	Essential oil, flavonoids, sulphur-containing compounds, phenylpropanoid derivatives, indole alkaloids, sterol glucosides, glucosinolates, phenanthrene, isothiocyanate anthocyanins	<i>Spodoptera littoralis</i> , <i>Escherichia coli</i> , <i>Klebsiella pneumonia</i> , <i>Salmonella paratyphi</i> , <i>Pseudomonas aeruginosa</i> , <i>Staphylococcus aureus</i>
<i>Bombax ceiba</i> Linn. ¹⁹³⁻¹⁹⁵	Shimul	Malvaceae	Flowers, young roots, leaves, gums, barks	Antibacterial, wound healing, tissue and bone regenerating properties, improves bowel movement, anti-diarrheal, aphrodisiac properties	Mangiferin, quercetin, shamimin, shamimoside, β -sitosterol, taraxeryl acetate, lupeol, simalin A, simalin B, shamimicin, bombaxalones A-D, bombaxquinone B, bombamalloside, bombasin	Bacteria, fungi, yeasts
<i>Cassia fistula</i> Linn. ¹⁹⁶⁻¹⁹⁷	Sondal	Caesalpiniaceae	Fruits, flowers, bark, roots	Anti-microbial, insecticidal, anti-neoplastic properties, anti-inflammatory, hepatoprotective, and hypoglycemic properties	Tannins, terpenoids, alkaloids, flavonoids, glycosides, anthraquinones, flavan-3-ol derivatives	Infectious bovine rhinotracheitis (IBR) virus
<i>Areca catechu</i> Linn. ¹⁹⁸⁻²⁰¹	Supari	Arecaceae	Nuts, buds, shoots, leaves	Antiparasitic, antibacterial, antiviral, improve oral hygiene and motility of food	Procyanidin, areca tannin B ₁ , tannins, terpenoids, alkaloids, flavonoids, fatty acids	Human immunodeficiency virus (HIV), gram-positive bacteria
<i>Cinnamomum tamala</i> Buch. Ham. ²⁰²⁻²⁰⁴	Tajpata	Lauraceae	Leaves, bark	Carminative, used in colic and diarrheal conditions	Reducing sugar, glycoside, tannins, steroid, amino acids, alkaloids, essential oils	Bacteria, fungi
<i>Ocimum sanctum</i> Linn. ²⁰⁵⁻²⁰⁸	Tulsi	Lamiaceae	Leaves	Rejuvenating and vitalizing properties, antiseptic, anti-allergic, anti-cancer effects	Oleanoic acid, rosmarinic acid, B-elemene, eugenol, limonene, carvacrol, germacrene	Newcastle disease virus (NDV), influenza virus (H ₃ N ₂)
<i>Solanum nigrum</i> Linn. ²⁰⁹⁻²¹¹	Tut begun	Solanaceae	Fruits, leaves	Antioxidant, anti-cancer, anti-inflammatory, diuretic, hepatoprotective, anti-bacterial, and anti-convulsant properties	Alkaloids, glycoprotein, polyphenolic components including catechin, gallic acid, epicatechin, caffeic acid, naringenin	Hepatitis C virus (HCV)
<i>Mucuna pruriata</i> Hook. / <i>Mucuna pruriens</i> Baker. ²¹²⁻²¹⁴	Ulkushi	Fabaceae	Seeds	Cholestrolemic, anti-parkinson, anti-diabetic, aphrodisiac, anti-inflammatory, anti-bacterial properties	Alkaloids, flavonoids, tannins, phenols	Bacteria, fungi, yeasts

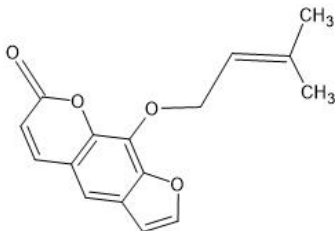
Isolated pure chemical compounds responsible for specific antiviral actions. Many investigations had been conducted to isolate pure chemical compounds from different medicinal plants and their potential

antiviral activities against various viruses were explored. A few chemical compounds found in some of these medicinal plants and their anti-viral activities are shown in table 3.

Table 3. The chemical structures of a variety of substances obtained from various medicinal plants.

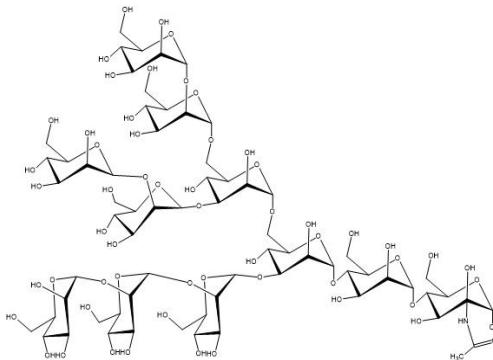
Active compound	Chemical structure	Anti-viral activity
Allicin ²¹⁵ (<i>Zingiber officinale</i> Roxb.)		Allicin is an influenza cytokine that has anti-viral activity against the Influenza virus (H1N1) ¹¹
Mangiferin ²¹⁶ (<i>Mangifera indica</i>)		Mangiferin is a tetrahydroxy pyrrolidone saponin extracted from <i>Mangifera indica</i> that possesses anti-viral activity against HSV-2 ¹⁶
1,2,4,6-Tetra-O-galloyl-β-D-glucose ²¹⁷ (1246TG G) (<i>Phyllanthus embelica</i>)		The polyphenolic compound 1246TGG suppresses the replication cycle of HSV-1 and HSV-2 ²⁰
Withafarin A ²¹⁸ (<i>Withania somnifera</i> Dunal.)		Withafarin A, as a neuraminidase inhibitor, has anti-viral efficacy against H ₁ N ₁ influenza neuraminidase (NA) and can limit NA activity by inhibiting the production of progeny viruses ²¹⁹
Curcuminoids ²²⁰ (<i>Curcuma longa</i> Linn.)		Curcuminoids derived from methanolic extracts of <i>Curcuma longa</i> were found to have significant inhibitory activity on the neuraminidases of several influenza virus variants such as H ₁ N ₁ and H ₉ N ₂ , when tested ⁸⁴

Marmelide⁵⁰
(*Aegle marmelos*
Corr.)



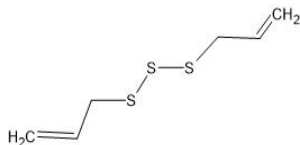
Marmelide is a pure compound that is the most effective virucidal agent against human coxsackievirus B₁-B₆.⁴⁸

Cyanovirin-N
(CVN)²²¹
(*Ceratopteris*
thalictroides)



Cyanovirin-N (CVN) isolated from *Ceratopteris thalictroides* is an anti-HIV protein.¹²²

Allitridin²²²
(*Allium sativum*
Linn.)



Allitridin, a diallyl trisulfide extracted from *A. sativum*, can suppress human cytomegalovirus (HCMV), IEA expression surprisingly *in vitro*, which is possibly a crucial mechanism in the anti-HCMV function of allitridine.¹⁷⁴

Recommendations. Hippocrates cured every disease with herbs and the art of healing from the magical-religious conceptions of the past freed by him. The abundance and potency of indigenous plants influenced him to acquire a strong belief in the healing properties of Mother Nature, which he used further for his medical studies. We, therefore, observe that nature is the major source of the drugs. In this review, we found that *Terminalia chebula* Retz, Haritaki in Bengali, contained different bioactive components which were effective in treatment of SARS COV-2 infection, hence, this plant showed potential antiviral activity against COVID-19.

CONCLUSION

In this paper we studied 62 Bangladeshi medicinal plants for the antiviral activity and found that 45 plants possess the activity. It has been acknowledged from ancient times that medicinal plants have established anti-viral efficacy against different viruses and in particular, against viral infections with characteristics similar to COVID-19.

Therefore, further investigation for isolation of the active constituents from the selective medicinal plants, elucidation of the biochemical mechanism of trafficking inhibition and *in vitro*, *in vivo* & *in silico* analyses are warranted. This appears to be a potential technique for decreasing the infectivity of corona virus in a worldwide situation and as a result, reducing the morbidity and mortality that has occurred due to COVID-19.

Consent for publication. All the authors who contributed to completing this review article have consent for the publication of this article. With consent, every author gives permission to publish this article and all authors guarantee that the information used in this article has not been previously published anywhere. It is to be noted that any one or more of the authors will not withdraw the consent later.

Author contributions. MSA is the one who came up with the original concept and laid out the general layout of the study. The first draft of the manuscript was written by AF, FA and TA. The illustrations in the manuscript have been drawn by AF. The literature survey was carried out by SK,

AAC, JAC and SMB. All authors improved the manuscript through mutual discussion by several offline and online meetings. Before submitting the final document, all authors have gone through it thoroughly and given their approval for the final version.

Funding. This research was carried out entirely based on self-funding.

Conflicts of interest. There are no conflicts of interest declared by the authors of the paper.

ACKNOWLEDGMENTS

In general and especially, we are grateful to the Department of Pharmaceutical Chemistry for permitting us to use their computer facilities in the Molecular Pharmacology and Herbal Drug Research Laboratory, which was established with a grant from the Higher Education Quality Enhancement Project (HEQEP), AIF, Round-III, Window-2, CP-3258, and the University Grants Commission (UGC) of Bangladesh.

REFERENCES

- Vaijwade, S. 2014. Screening of antiviral compounds from plants-a review. *J. Pharm. Res.* **8**, 1050-1058.
- Ben-Shabat, S., Yarmolinsky, L., Porat, D. and Dahan, A. 2020. Antiviral effect of phytochemicals from medicinal plants: Applications and drug delivery strategies. *Drug Deliv. Transl. Res.* **10**, 354-367.
- Haque, E., Karim, A., Chowdhury, J.A., Rezwani, R., Akter, T., Tahsin, R., Chowdhury, A.A., Kabir, S. and Shah, M.A. 2021. Can *Terminalia Chebula* (Haritaki) Stop COVID-19. *Eur. J. Pharm. Med. Res.* **8**, 115-119.
- Hossain, M.F., Hasana, S., Mamun, A.A., Uddin, M.S., Wahed, M.I.I. and Sarker, S. 2020. COVID-19 Outbreak: Pathogenesis, Current Therapies, and Potentials for Future Management. *Front Pharmacol.* **11**, 1-25.
- Ryan, K.J. and Ray, C.G. 2004. Medical microbiology. McGraw Hill., 4.
- De Clercq, E. 2007. Viruses and viral diseases. *Comprehensive Medicinal Chemistry II*, 253.
- Gamage, K., Dissanayake, C., Angoda, W., Chandrasiri, W. L. and Liyanage, R.P. 2020. A Review on Medicinal Uses of *Zingiber officinale* (Ginger). *Int. J. Health Sci Res.* **10**, 142-148.
- Chang, J.S., Wang, K.C., Yeh, C.F., Shieh, D.E. and Chiang, L.C. 2013. Fresh ginger (*Zingiber officinale*) has anti-viral activity against human respiratory syncytial virus in human respiratory tract cell lines. *J Ethnopharmacol.* **145**, 146-151.
- El-Wahab, A., Eladawi, H. and El Demellawy, M. 2009. In vitro study of the antiviral activity of *Zingiber officinale*. *Planta Med.* **75**, 75 - PF7, DOI: 10.1055/s-0029-1234649
- Abdel-Moneim, A., Morsy, B.M., Mahmoud, A.M., Abo-Seif, M.A. and Zanaty, M.I. 2013. Beneficial therapeutic effects of *Nigella sativa* and/or *Zingiber officinale* in HCV patients in Egypt. *EXCLI J.* **12**, 943-955.
- Sahoo, M., Jena, L., Rath, S.N. and Kumar, S. 2016. Identification of suitable natural inhibitor against influenza A (H₁N₁) neuraminidase protein by molecular docking. *Genom Inform.* **14**, 96-103.
- Koch, C., Reichling, J., Schneele, J. and Schnitzler, P. 2008. Inhibitory effect of essential oils against herpes simplex virus type 2. *Phytomed.* **15**, 71-80.
- Khairnar, A.K., Bhamare, S.R. and Bhamare, H.P. 2012. *Calotropis procera*: an ethnopharmacological update. *Adv. Res. Pharmaceut. Biol.* **2**, 142-156.
- Saher, U., Javed, A., Ashraf, M., Altaf, I. and Ghafoor, A. 2018. Evaluation of antiviral and cytotoxic activity of *Calotropis procera* against foot and mouth disease virus. *Int. J. Sci. Eng. Res.* **9**, 236-253.
- Shah, K.A., Patel, M.B., Patel, R.J. and Parmar, P.K. 2010. *Mangifera indica* (mango). *Pharmacogn Rev.* **4**, 42-48.
- Zhu, X.M., Song, J.X., Huang, Z.Z., Wu, Y.M. and Yu, M.J. 1993. Antiviral activity of mangiferin against herpes simplex virus type 2 in vitro. *Zhongguo Yao Li Xue Bao.* **14**, 452-454.
- Rechenchoski, D.Z., Samensari, N.L., Faccin-Galhardi, L.C., de Almeida, R.R., Cunha, A.P., Ricardo, N.M., Nozawa, C. and Linhares, R.E. 2019. The combination of dimorphandra gardneriana galactomannan and mangiferin inhibits herpes simplex and poliovirus. *Curr. Pharmaceut. Biotech.* **20**, 215-221.
- Gaire, B.P. and Subedi, L. 2014. Phytochemistry, pharmacology and medicinal properties of *Phyllanthus emblica* Linn. *Chin. J. Integr. Med.* **20**, 1-8.
- Estari, M., Venkanna, L., Sriprya, D. and Lalitha, R. 2012. Human Immunodeficiency Virus (HIV-1) reverse transcriptase inhibitory activity of *Phyllanthus emblica* plant extract. *Biol. Med.* **4**, 178-182.
- Xiang, Y., Pei, Y., Qu, C., Lai, Z., Ren, Z. and Yang, K. 2011. In vitro anti-herpes simplex virus activity of 1,2,4,6-tetra-O-galloyl- β -D-glucose from *Phyllanthus emblica* L. (Euphorbiaceae). *Phytotherapy Res.* **25**, 975-982.
- Mishra, N.N., Kesharwani, A., Agarwal, A., Polachira, S.K., Nair, R. and Gupta, S.O.K. 2018. Herbal gel formulation developed for anti-human immunodeficiency virus (HIV)-1 activity also inhibits in vitro HSV-2 infection. *Viruses.* **10**, 580-587.
- Al-snafi, A.E. 2016. Pharmacological importance of *Clitoria ternatea*- A review. *J. Pharm.* **6**, 68-83.
- Kamilla, L., Mnsor, S.M., Ramanathan, S. and Sasidharan, S. 2009. Antimicrobial activity of *Clitoria ternatea* (L.) extracts. *Pharmacologyonline.* **1**, 731-738.

24. Jain, S., Yadav, P.P., Gill, V., Vasudeva, N. and Singla, N. 2009. *Terminalia arjuna* a sacred medicinal plant: Phytochemical Pharmacological profile. *Phytochem Rev.* **8**, 491-502.
25. Cheng, H.Y., Lin, C.C. and Lin, T.C. 2002. Antiherpes simplex virus type 2 activity of casuarinin from the bark of *Terminalia arjuna* Linn. *Antiviral Res.* **55**, 447-455.
26. Khan, Z.H., Faruquee, H.M. and Shaik, M.M. 2013. Phytochemistry and pharmacological potential of *Terminalia arjuna* L. *Med. Plant Res.* **3**, 70-77.
27. Pradhan, P., Joseph, L., Gupta, V., Chulet, R., Arya, H., Verma, R. and Bajpai, A. 2009. *Saraca asoca* (Ashoka): a review. *J. Chem Pharmaceut. Res.* **1**, 62-71.
28. Chakraborty, R., Sen, S., Deka, M.K., Rekha, B. and Sachan, S. 2014. Anti-microbial evaluation of *Saraca indica* leaves extracts by disk diffusion method. *J. Pharm. Chem. Biol. Sci.* **1**, 1-5.
29. Dar, N.J., Hamid, A. and Ahmad, M. 2015. Pharmacologic overview of *Withania somnifera*, the Indian Ginseng. *Cell Mole Life Sci.* **72**, 4445-4460.
30. Kambizi, L., Goosen, B.M., Taylor, M. and Afolayan, A. 2007. Anti-viral effects of aqueous extracts of *Aloe ferox* and *Withania somnifera* on herpes simplex virus type 1 in cell culture. *S. Afr. J. Sci.* **103**, 359-360.
31. Cai, Z., Zhang, G., Tang, B., Liu, Y., Fu, X. and Zhang, X. 2015. Promising anti-influenza properties of active constituent of *withania somnifera* ayurvedic herb in targeting neuraminidase of h1n1 influenza: computational study. *Cell Biochem. Biophys.* **72**, 727-739.
32. Kumari, S., Krishna, M.J., Joshi, A.B., Gurav, S., Bhandarkar, A.V. and Agarwal, A. 2017. A pharmacognostic, phytochemical and pharmacological review of *Terminalia bellerica*. *J. Pharmacog Phytochem.* **6**, 368-376.
33. Valsaraj, R., Pushpangadan, P., Smitt, U.W., Adersen, A., Christensen, S.B. and Sittie, A. 1997. New anti-HIV-1, antimalarial, and antifungal compounds from *Terminalia bellerica*. *J. Nat. Prod.* **60**, 739-742.
34. Chanda, S., Biswas, S.M. and Sarkar, P.K. 2020. Phytochemicals and antiviral properties of five dominant medicinal plant Phytochemicals and antiviral properties of five dominant medicinal plant species in Bankura district, West Bengal: An overview. *J. Pharmacogn. Phytochem.* **9**, 1420-1427.
35. Shabbir, A., Khan, M.M., Ahmad, B., Sadiq, Y., Jaleel, H. and Uddin, M. 2019. *Vetiveria zizanioides* (L.) Nash: A Magic Bullet to Attenuate the Prevailing Health Hazards. *Plant and Human Health.* **2**, 99-120.
36. Durge, A.A. and Moon, U.R. 2021. *Vetiveria zizanioides* Nash-a wonder plant: an industrial perspective. *J. Adv. Sci. Res. ICITNAS*, 01-06.
37. Lavanya, P., Ramaiah, S. and Anbarasu, A. 2016. Ethyl 4-(4-methylphenyl)-4-pentenoate from *Vetiveria zizanioides* inhibits dengue NS2B-NS3 protease and prevents viral assembly: a computational molecular dynamics and docking study. *Cell Biochem. Biophys.* **74**, 337-351.
38. Khatun, F., Mahfuz-E-Alam, M., Tithi, N.S., Nasrin, N. and Asaduzzaman, M. 2015. Evaluation of phytochemical, antioxidant, anthelmintic and antimicrobial properties of *Crataeva nurvala* buch. Ham. Leaves. *Int. J. Pharma. Sci. Res.* **6**, 1422-1429.
39. Ali, S.G., Ansari, M.A., Khan, H.M., Jalal, M, Mahdi, A.A. and Cameotra, S.S. 2017. *Crataeva nurvala* nanoparticles inhibit virulence factors and biofilm formation in clinical isolates of *Pseudomonas aeruginosa*. *J. Basic Microbiol.* **57**, 193-203.
40. Singh, S.K., Patel J.R., Dangi, A., Bachle, D. and Kataria, R.K. 2017. A complete over review on *Adhatoda vasica* a traditional medicinal plants. *J Med plants stud.* **5**, 175-80.
41. Singh, K.P., Upadhyay, R. and Prasad, A.K. 2010. Screening of *Adhatoda vasica* Nees as a Putative HIV-protease Inhibitor. *J. Phytol.* **2**, 78-82.
42. Shoaib, A. 2021. A systematic ethnobotanical review of *Adhatoda vasica* (L.), Nees. *Cell Mol. Biol.* **67**, 248-263.
43. Chavan, R. and Chowdhary, A. 2014. *In vitro* inhibitory activity of *Justicia adhatoda* extracts against influenza virus infection and hemagglutination. *Int. J. Pharm. Sci. Rev. Res.* **25**, 231-236.
44. Al-Snafi, A. 2015. The chemical constituents and pharmacological effects of *Chenopodium album*-an overview. *Int. J. Pharmacol. Screen Met.* **5**, 10-17.
45. Dutt, S., Yadav, O.P., Kapoor, H.C. and Lodha, M.L. 2004. Possible mechanism of action of antiviral proteins from the leaves of *Chenopodium album* L. *Indian J. Biochem. Biophys.* **41**, 29-33.
46. Rajan, S., Gokila, M., Jency, P., Brindha, P. and Sujatha, R.K. 2011. Antioxidant and phytochemical properties of *Aegle marmelos* fruit pulp. *Int. J. Curr. Pharm. Res.* **3**, 65-70.
47. Gupta, D., John, P., Pankaj, K., Kaushik, R. and Yadav, R. 2011. Pharmacological Review of *Aegle Marmelos* Corr. Fruits. *Int. J. Pharm Sci. Res.* **2**, 2278-2285.
48. Maity, P., Hansda, D., Bandyopadhyay, U. and Mishra, D.K. 2009. Biological activities of crude extracts and chemical constituents of bael, *Aegle marmelos* (L.) Corr. *Indian J. Exp. Biol.* **47**, 849-861.
49. Badam, L., Bedekar, S.S., Sonawane, K.B. and Joshi, S.P. 2002. *In vitro* antiviral activity of bael (*Aegle marmelos* Corr) upon human coxsackieviruses B1-B6. *J. Commun. Diss.* **34**, 88-99.
50. Lamia, S.S., Shimo, M.S., Rashed, S.S., Prima, A.A., Mony, A.T. and Dash, P.R. Phytochemistry and pharmacological properties of *Aegle marmelos* L (Rutaceae): A review. *Phytochem.* **3**, 45-54.
51. Kadri, A., Gharsallah, N., Damak, M. and Gdoura, R. 2011. Chemical composition and in vitro antioxidant properties of essential oil of *Ricinus communis* L. *J. Med. Plants Res.* **5**, 1466-1470.
52. Hamidi, J.A., Ismaili, N.H., Ahmadi, F.B. and Lajisi, N.H. 1996. Antiviral and cytotoxic activities of some plants used in Malaysian indigenous medicine. *Pertanika J. Trop. Agr. Sci.* **19**, 129-136.

53. Murade, V., Hase, D., Deshmukh, K. and Pansambal, S. 2014. A comprehensive review of phytopharmacology of *Ricinus Communis* (Linn.). *Int. J. Phytopharm.* **5**, 328-334.
54. Bharti, R., Chopra, B.S.O, Raut, S. and Khatri, N. 2021. *Pueraria tuberosa*: a review on traditional uses, pharmacology, and phytochemistry. *Front Pharmacol.* **11**, 1-19.
55. Sadguna, V., Sarikha, K., Komuraiah, T.R. and Mustafa, M. 2015. Anti-microbial Activity of *Pueraria tuberosa* DC, an Economically and Medicinally Important Plant. *Int. J. Curr. Microbiol. App. Sci.* **4**, 152-159.
56. Paithankar, V.V., Raut, K.S., Charde, R.M. and Vyas, J.V. 2011. *Phyllanthus niruri*: A magic herb. *Research in Pharmacy.* **1**, 1-9.
57. Wahyuni, T.S., Azmi, D., Permanasari, A.A., Adianti, M., Tumewu, L. and Widiandani, T. 2019. Anti-viral activity of *Phyllanthus niruri* against hepatitis C virus. *Malaysian App. Biol.* **48**, 105-111.
58. Ogata, T., Higuchi, H., Mochida, S., Matsumoto, H., Kato, A., Endo, T., Kaji, A. and Kaji, H. 1992. HIV-1 reverse transcriptase inhibitor from *Phyllanthus niruri*. *AIDS Res. Hum. Retrovir.* **8**, 1937-1944.
59. Venkateswaran, P.S., Millman, I. and Blumberg, B.S. 1987. Effects of an extract from *Phyllanthus niruri* on hepatitis B and woodchuck hepatitis viruses: *In vitro* and *in vivo* studies. *Proc. Nat. Acad. Sci. USA.* **84**, 274-280.
60. Sampathkumar, P., Dheeba, B., Vidhyasagar, V., Arulprakash, T. and Vinothkannan, R. 2008. Potential antimicrobial activity of various extracts of *Bacopa monnieri* (Linn.). *Int. J. Pharmacol.* **4**, 230-232.
61. Verma, P.K., Raina, R., Agarwal, S. and Kaur, H. 2018. Phytochemical ingredients and Pharmacological potential of *Calendula officinalis* Linn. *Pharmaceut. Biomed. Res.* **4**, 1-7.
62. Kalvatchev, Z., Walder, R. and Garzaro, D. 1997. Anti-HIV activity of extracts from *Calendula officinalis* flowers. *Biomed Pharmacotherapy.* **51**, 176-180.
63. Sayyed, A., Shah, M. and Aqib Sayyed, C. 2014. Phytochemistry, pharmacological and traditional uses of *Datura stramonium* L. review. *J. Pharmacogn. Phytochem.* **2**, 123-125.
64. Appenzeller, O. 1997. Primer on the Autonomic Nervous System. *Trends in Neurosciences.* **1**, 49-50.
65. Yamazaki, Z. and Tagaya, I. 1980. Antiviral effects of atropine and caffeine. *J. General Virology.* **50**, 429-431.
66. Roy, S., Mukherjee, S., Pawar, S. and Chowdhary, A. 2016. Evaluation of *in vitro* antiviral activity of *Datura metel* Linn. against rabies virus. *Pharmacog. Res.* **8**, 265-269.
67. Vimalanathan, S., Ignacimuthu, S. and Hudson, J.B. 2009. Medicinal plants of Tamil Nadu (Southern India) are a rich source of antiviral activities. *Pharm. Biol.* **47**, 422-429.
68. Kundu, S., Salma, U., Sutradhar, M. and Mandal, N. 2018. An update on the medicinal uses, phytochemistry and pharmacology of *Leucas aspera*, a medicinally important species. *Int. Agr. Innov. Res.* **6**, 39-44.
69. Al-Snafi, P.D.A.E. 2016. Chemical constituents and pharmacological effects of *Cynodon dactylon*- A Review. *IOSR J. Pharm. (IOSRPHR).* **6**, 17-31.
70. Shendye, N.V. and Gurav, S.S. *Cynodon dactylon*: A systemic review of pharmacognosy, Phytochemistry and pharmacology. *Int. J. Pharm. Pharm. Sci.* **6**, 7-12.
71. Balasubramanian, G., Sarathi, M., Venkatesan, C., Thomas, J. and Sahul Hameed, A.S. 2008. Oral administration of antiviral plant extract of *Cynodon dactylon* on a large scale production against White spot syndrome virus (WSSV) in *Penaeus monodon*. *Aquaculture.* **279**, 2-5.
72. Pringproa, K., Khonghiran, O., Kunanoppadol, S., Potha, T. and Chuammitri, P. 2014. *In vitro* virucidal and virustatic properties of the crude extract of *cynodon dactylon* against porcine reproductive and respiratory syndrome virus. *Vet. Med. Int.* **2014**, 1-5.
73. Murali, K.S., Sivasubramanian, S., Vincent, S., Murugan, S.B., Giridaran, B. and Dinesh, S. 2015. Anti-chikungunya activity of luteolin and apigenin rich fraction from *Cynodon dactylon*. *Asian Pac. J. Trop. Med.* **8**, 352-358.
74. Warriar, R.R., Priya, S.M. and Kalaiselvi, R. 2021. *Gmelina arborea*– an indigenous timber species of India with high medicinal value: A review on its pharmacology, pharmacognosy and phytochemistry. *J. Ethnopharmacol.* **267**, 1-4.
75. Dighe, V.V., Pathak, G.M., Tulpule, K.M. and Gokarn, V.N. 2007. HPTLC method for quantification of apigenin in the dried root powder of *Gmelina arborea* Linn. *J. Planar. Chromatog. Mod. TLC.* **20**, 179-182.
76. Ajah, O., Omodamiro, O.D., Jimoh, M.A. and Ewa-ibe, C. 2020. Evaluation of anti-diarrhea and antimicrobial activities of methanol extract leaves of *Gmelina arborea*. *J. Med. Botany.* **4**, 20-23.
77. Sharma, P., Dwivedee, B.P., Bisht, D., Dash, A.K. and Kumar, D. 2019. The chemical constituents and diverse pharmacological importance of *Tinospora cordifolia*. *Heliyon.* **5**, 1-8.
78. Maddi, R., Kandula, V.L., Vallepu, B., Navuluri, H. and Kollu, H. 2018. Preliminary Phytochemical Analysis and *In vitro* Antiviral Activity of Ethanolic extract of Whole plant of *Tinospora cordifolia* (Thunb.) Miers against Hepatitis-A Virus. *Int. J. Sci. Res. Biol. Sci.* **5**, 51-55.
79. Krup, V., Prakash, H.L. and Harini, A. 2013. Pharmacological Activities of Turmeric (*Curcuma longa* linn): A Review. *J. Homeo. Ayurv Med.* **2**, 133-137.
80. Rivera-Espinoza, Y. and Muriel, P. 2009. Pharmacological actions of curcumin in liver diseases or damage. *Liver Int.* **29**, 1457-1466.
81. Kim, H.J., Yoo, H.S., Kim, J.C., Park, C.S., Choi, M.S. and Kim, M. 2009. Antiviral effect of *Curcuma longa* Linn extract against hepatitis B virus replication. *J. Ethnopharmacol.* **124**, 189-196.
82. Tamam, S.M., Madbouly, H.M. and Amin, F. 2010. Antiviral activity of *Curcuma longa* against Newcastle disease virus (*in vitro* and *in vivo* studies). *J. Vet. Med. Res.* **20**, 290-295.

83. Ichsyani, M., Ridhanya, A., Risanti, M., Desti, H., Ceria, R. and Putri, D.H. 2017. Antiviral effects of *Curcuma longa* L. against dengue virus *in vitro* and *in vivo*. *IOP Conf Ser: Earth Environ Sci.* **101**, 0-10.
84. Dao, T.T., Nguyen, P.H., Won, H.K., Kim, E.H., Park, J. and Won, B.Y. 2012. Curcuminoids from *Curcuma longa* and their inhibitory activities on influenza A neuraminidases. *Food Chem.* **134**, 21-28.
85. Niranjan, A. and Prakash, D. 2008. Chemical constituents and biological activities of turmeric (*Curcuma longa* L.)-a review. *J. Food Sci. Tech.* **45**, 109-116.
86. Lee, D., Boo, K.H., Woo, J.K., Duan, F., Lee, K.H. and Kwon, T.K. Anti-bacterial and anti-viral activities of extracts from *Terminalia chebula* barks. *J. Appl. Biol. Chem.* **54**, 295-298.
87. Kesharwani, A., Polachira, S.K., Nair, R., Agarwal, A., Mishra, N.N. and Gupta, S.K. 2017. Anti-HSV-2 activity of *Terminalia chebula* Retz extract and its constituents, chebulagic and chebulinic acids. *BMC Complement Altern Med.* **17**, 110-121.
88. Kim, T.G., Kang, S.Y., Jung, K.K., Kang, J.H., Lee, E. and Han, H.M. 2001. Antiviral activities of extracts isolated from *Terminalia chebula* Retz., *Sanguisorba officinalis* L., *Rubus coreanus* Miq. and *Rheum palmatum* L. against hepatitis B virus. *Phytotherapy Res.* **15**, 718-720.
89. Nigam, M., Mishra, A.P., Adhikari-Devkota, A., Dirar, A.I., Hassan, M.M., Adhikari, A., Belwal, T. and Devkota, H.P. 2020. Fruits of *Terminalia chebula* Retz.: A review on traditional uses, bioactive chemical constituents and pharmacological activities. *Phytotherapy Res.* **34**, 2518-2533.
90. Ziai, S., Hamkar, R., Monavari, H., Norooz-Babaei, Z. and Adibi, L. 2007. Antiviral Effect Assay of Twenty Five Species of Various Medicinal Plants Families in Iran. *J. Med. Plants.* **6**, 1-9.
91. Adhikari, P.P. and Paul, S.B. 2018. Medicinally important plant *Cleome gynandra*: a phytochemical and pharmacological explanation. *Asian J. Pharm. Clin. Res.* **11**, 21-29.
92. Sridhar, N., Surya Kiran, B.V.V.S., Tharaka Sasidhar, D. and Kanthal, L.K. 2014. *In vitro* antimicrobial screening of methanolic extracts of *Cleome chelidonii* and *Cleome gynandra*. *Bangladesh J. Pharmacol.* **9**, 161-166.
93. Imanirampa, L. and Alele, P.E. 2016. Antifungal activity of *Cleome gynandra* L. aerial parts for topical treatment of *Tinea capitis*: An *in vitro* evaluation. *BMC Complement Altern Med.* **16**, 194-202.
94. Ganesh, S., Muthusamy, S. and Jaganathan, V. 2018. Preliminary Phytochemical screening, Anti-bacterial and Thrombolytic activity of *Cleome gynandra* aqueous extract. *Int. J. Adv. Res. Biol. Sci.* **5**, 30-36.
95. Unani, A.N., Umer, K.H., Zeenat, F., Ahmad, W. and Vakil, A. 2017. Therapeutics, phytochemistry and pharmacology of Ikhlilul Malik (*Astragalus hamosus* Linn): A natural Unani Remedy. *Intern. J. Herb. Med.* **5**, 1-5.
96. Kumbhar, S.T., Patil, S.P. and Une, H.D. 2018. Phytochemical analysis of *Canna indica* L. roots and rhizomes extract. *Biochem Biophys Rep.* **16**, 50-55.
97. Thepouyporn, A., Yoosook, C., Chuakul, W., Thirapanmethee, K., Napaswad, C. and Wiwat, C. 2012. Purification and characterization of anti-HIV-1 protein from *Canna indica* L. leaves. *Southeast Asian J Trop Med Pub Health.* **43**, 1153-1159.
98. Dalli, M., Bekkouch, O., Azizi, S.E., Azghar, A., Gseyra, N. and Kim, B. 2021. *Nigella sativa* L. Phytochemistry and Pharmacological Activities: A Review (2019–2021). *Biomolecules.* **12**, 20-57.
99. Salem, M.L. 2005. Immunomodulatory and therapeutic properties of the *Nigella sativa* L. seed. *Int Immunopharmacol.* **5**, 1749-1770.
100. Ahmad, A., Husain, A., Mujeeb, M., Khan, S.A., Najmi, A.K. and Siddique, N.A, 2013. A review on therapeutic potential of *Nigella sativa*: A miracle herb. *Asian Pac. J. Trop. Biomed.* **3**, 337-352.
101. Barakat, A.B., Shoman, S.A., Dina, N. and Alfarouk, O.R. 2010. Antiviral activity and mode of action of *Dianthus caryophyllus* L. and *Lupinus termis* L. seed extracts against *in vitro* herpes simplex and hepatitis A viruses infection. *J. Microbiol. Antimicrob.* **2**, 23-29.
102. Dorra, N., El-Berrawy, M., Sallam, S. and Mahmoud, R. 2019. Evaluation of Antiviral and Antioxidant Activity of Selected Herbal Extracts. *J. High Inst. of Pub. Health.* **49**, 36-40.
103. Ulasli, M., Gurses, S.A., Bayraktar, R., Yumrutas, O., Oztuzcu, S. and Igcı, M. 2014. The effects of *Nigella sativa* (Ns), *Anthemis hyalina* (Ah) and *Citrus sinensis* (Cs) extracts on the replication of coronavirus and the expression of TRP genes family. *Mol. Biol. Rep.* **41**, 1703-1711.
104. Barakat, E.M.F., El Wakeel, L.M. and Hagag, R.S. 2013. Effects of *Nigella sativa* on outcome of hepatitis C in Egypt. *World J. Gastroenterol.* **19**, 2529-2536.
105. Mali, R.G., Mahajan, S. and Mehta, A. 2007. Rakta Kanchan (*Bauhinia variegata*): Chemistry, Traditional and Medicinal uses- A Review. *Pharmacogn. Rev.* **1**, 314-319.
106. Shaheen, M., El-Gamal, M., Mousa, A., Mostafa, S. and El-Esnawy, N. 2014. Antiviral activity of *Bauhinia variegata* extracts against rotavirus *in vitro*. *Curr. Sci. Int.* **3**, 172-178.
107. Parmar, S., Gangwal, A. and Sheth, N. 2010. *Solanum xanthocarpum* (yellow berried night shade): a review. *Der. Pharm. Lett.* **2**, 373-383.
108. Salar, R. and Devi, C. 2009. Evaluation of antimicrobial potential of different extracts of *Solanum xanthocarpum* Schrad. and Wendl. *Afr. J. Microbiol. Res.* **3**, 97-100.
109. Dey, P. and Chaudhuri, T.K. 2014. Pharmacological aspects of *Nerium indicum* Mill: A comprehensive review. *Pharmacogn Rev.* **8**, 156-162.
110. Singh, S., Shenoy, S., Nehete, P.N., Yang, P., Nehete, B. and Fontenot, D. 2013. *Nerium oleander* derived cardiac glycoside oleandrin is a novel inhibitor of HIV infectivity. *Fitoterapia.* **84**, 32-39.

111. Rajbhandari, M., Wegner, U., Jülich, M., Schoepke, T. and Mentel, R. 2001. Screening of Nepalese medicinal plants for antiviral activity. *J. Ethnopharmacol.* **74**, 251-255.
112. Sood, P. and Shri, R. 2018. A review on ethnomedicinal, phytochemical and pharmacological aspects of *Myrica esculenta*. *Indian J. Pharmaceut. Sci.* **80**, 2-13.
113. Agnihotri, S., Wakode, S. and Ali, M. 2012. Essential oil of *Myrica esculenta* Buch. Ham.: Composition, antimicrobial and topical anti-inflammatory activities. *Nat. Prod. Res.* **26**, 2266-2269.
114. Saklani, S.A., Chandra, S.U., Mishra, A.P.O and Badoni, P.P. 2012. Nutritional evaluation, antimicrobial activity and phytochemical screening of wild edible fruit of *Myrica nagi* pulp. *Int. J. Pharm. Pharmaceut. Sci.* **4**, 407-411.
115. Shan, B., Cai, Y.Z., Brooks, J.D. and Corke, H. 2007. The *in vitro* antibacterial activity of dietary spice and medicinal herb extracts. *Int. J. Food Microbiol.* **117**, 112-119.
116. Mann, S., Satpathy, G. and Gupta, R.K. 2015. *In vitro* evaluation of bio-protective properties of underutilized *Myrica esculenta* Buch. Ham. ex D. Don fruit of Meghalaya. *Indian J. Nat. Prod. Resour.* **6**, 183-188.
117. Rashid, M., Shamsi, S., Zaman, R. and Ilahi, A. 2015. Kath (Acacia catechu): An Overarching Envelop of Traditional and Modern Update. *Int. J. Curr. Trends pharm. Res.* **3**, 1007-1012.
118. Li, X., Wang, H., Liu, C. and Chen, R. 2010. Chemical constituents of *Acacia catechu*. *Zhongguo Zhong yao Za zhi.* **35**, 1425-1427.
119. Gupta, A. and Chaphalkar, S.R. 2016. Cytotoxic and antiviral activity of *Acacia Catechu* on human peripheral blood mononuclear cells. *Indonesian J. Pharm.* **27**, 111-116.
120. Smitha, V., Priyadarshana, M., Girija, M. and Vadivel, V. 2022. Green synthesis of silver nanoparticles from ceratopteris thalictroides: synthesis and characterization. *Int. J. Biol. Pharm. Allied sci.* **1**, 2520-2530.
121. Smitha, V. and Vadivel, V. 2019. Phytochemical screening for active compounds in *Ceratopteris thalictroides* (L.) Brogn. *J Pharmacog Phytochem.* **8**, 3556-3559.
122. Gustafson, K.R., Sowder, R.C., Henderson, L.E., Cardellina, J.H., McMahon, J.B. and Rajamani, U. 1997. Isolation, primary sequence determination, and disulfide bond structure of cyanovirin-N, an anti-HIV (Human Immunodeficiency Virus) protein from the cyanobacterium *Nostoc ellipsosporum*. *Biochem. Biophys Res. Commun.* **238**, 223-228.
123. Sun, J., Su, Y. and Wang, T. 2013. Expression, purification and identification of CtCVNH, a novel anti-HIV (human immunodeficiency virus) protein from *Ceratopteris thalictroides*. *Int. J. Mol. Sci.* **14**, 7506-7514.
124. Gopinath, G., Thirumal, M. and Kumar, P.R. 2020. *Holarrhena antidysenterica* Linn. – A Review. *Res. J. Pharm. Technol.* **13**, 2013-2019.
125. Sinha, S., Sharma, A., Reddy, P.H., Rathi, B., Prasad, N.V.S.R.K. and Vashishtha, A. 2013. Evaluation of phytochemical and pharmacological aspects of *Holarrhena antidysenterica* (Wall.): A comprehensive review. *J. Pharm. Res.* **6**, 488-492.
126. Ballal, M., Srujan, D., Bhat, K.K., Shirwaikar, A. and Shivananda, P.G. 2000. Antibacterial activity of *Holarrhena antidysenterica* [Kurchi] against the enteric pathogens. *Indian J. Pharmacol.* **32**, 392-393.
127. Parekh, J. and Chanda, S. 2008. In vitro antifungal activity of methanol extracts of some Indian medicinal plants against pathogenic yeast and moulds. *Afr. J. Biotechnol.* **7**, 4349-4353.
128. Asgarpanah, J. and Kazemivash, N. 2013. Phytochemistry, pharmacology and medicinal properties of *Carthamus tinctorius* L. *Chin. J. Integr. Med.* **19**, 153-159.
129. Zhang, L.L., Tian, K., Tang, Z.H., Chen, X.J., Bian, Z.X., Wang, Y.T. and Lu, J.J. 2016. Phytochemistry and Pharmacology of *Carthamus tinctorius* L. *American J. Chin. Med.* **44**, 197-226.
130. Zhang, H. and Ma, Z.F. 2018. Phytochemical and pharmacological properties of *Capparis spinosa* as a medicinal plant. *Nutrients.* **10**, 116-121.
131. Arena, A., Bisignano, G., Pavone, B., Tomaino, A., Bonina, F.P. and Saija, A. 2008. Antiviral and immunomodulatory effect of a lyophilized extract of *Capparis spinosa* l. buds. *Phytotherapy Res.* **22**, 313-317.
132. Al-snafi, A.E. 2015. Pharmacology and Medicinal Properties of *Caesalpinia crista*, an overview. *Int. J. Pharm.* **5**: 71-83.
133. Usha, P. and Sharma, M.C. 2012. Antiviral activity of lathakaranja (*Caesalpinia crista* L.) crude extracts on selected animal viruses. *Global J. Res. Med. Plants Indigen. Med.* **1**, 440-447.
134. Ahmad, H., Sehgal, S., Mishra, A. and Gupta, R. 2012. *Mimosa pudica* L. (Laajvanti): an overview. *Pharmacognosy Rev.* **6**, 115-124.
135. Malayan, J., Selvaraj, B., Warriar, A., Shanmugam, S., Mathayan, M. and Menon, T. 2013. Anti-mumps virus activity by extracts of *Mimosa pudica*, a unique Indian medicinal plant. *Indian J. Virol.* **24**, 166-173.
136. Dini, I. 2018. Spices and Herbs as Therapeutic Foods. Food Quality: Balancing Health and Disease. **13**, 433-469.
137. Sharma, V., Singh, P. and Rani, A. 2017. Antimicrobial activity of *Trigonella foenum-graecum* L. Fenugreek). *Eur. Exp. Biol.* **7**, 1-4.
138. Goyal, S., Gupta, N. and Chatterjee, S. 2016. Investigating therapeutic potential of *Trigonella foenum-graecum* L. as our defense mechanism against several human diseases. *J.Toxicol.* **2016**, 1-10.
139. Sayeed, R., Thakur, M. and Gani, A. 2020. *Celosia cristata* Linn. flowers as a new source of nutraceuticals- A study on nutritional composition, chemical characterization and *in-vitro* antioxidant capacity. *Heliyon.* **6**, 1-10.
140. Gholizadeh, A. and Kapoor, H. 2004. Modifications in the Purification Protocol of *Celosia Cristata* Antiviral Proteins Lead to Protein that Can Be N-Terminally Sequenced. *Protein Pept Lett.* **11**, 555-561.

141. Herrmann, F., Romero, M.R., Blazquez, A.G., Kaufmann, D., Ashour, M.L., Kahl, S., Marin, J.J.G., Efferth, T. and Wink, M. 2011. Diversity of Pharmacological Properties in Chinese and European Medicinal Plants: Cytotoxicity, Antiviral and Antitrypanosomal Screening of 82 Herbal Drugs. *Diversity*. **3**, 547-580.
142. Ju, Y. and Xiao, B. 2016. Chemical constituents of *Cyperus rotundus* L. and their inhibitory effects on uterine fibroids. *Afr. Health Sci*. **16**, 1000-1006.
143. Peerzada, A.M., Ali, H.H., Naeem, M., Latif, M., Bukhari, A.H. and Tanveer, A. 2015. *Cyperus rotundus* L.: Traditional uses, phytochemistry, and pharmacological activities. *J. Ethnopharmacol*. **174**, 540-560.
144. Parvez, M.K., Al-Dosari, M.S., Arbab, A.H. and Niyazi, S. 2019. The *in vitro* and *in vivo* anti-hepatotoxic, anti-hepatitis B virus and hepatic CYP450 modulating potential of *Cyperus rotundus*. *Saud. Pharmaceut. J*. **27**, 558-564.
145. Samra, R.M., Soliman, A.F., Zaki, A.A., El-Gendy, A.N., Hassan, M.A. and Zaghloul, A.M. 2020. Chemical Composition, Antiviral and Cytotoxic Activities of Essential Oil from *Cyperus rotundus* Growing in Egypt: Evidence from Chemometrics Analysis. *J. Essen. Oil-Bearing Plants*. **23**, 648-659.
146. Xu, H.B., Ma, Y.B., Huang, X.Y., Geng, C.A., Wang, H. and Zhao, Y. 2015. Bioactivity-guided isolation of anti-hepatitis B virus active sesquiterpenoids from the traditional Chinese medicine: Rhizomes of *Cyperus rotundus*. *J. Ethnopharmacol*. **171**, 131-140.
147. Vinoth, B., Manivasagerumal, R. and Rajaravindran, M. 2012. Phytochemical analysis and antibacterial activity of *Azadirachta indica* A. Juss. *Int J Res Plant Sci*. **2**, 50-55.
148. Hashmat, I., Azad, H. and Ahmed, A. 2012. Neem (*Azadirachta indica* A. Juss) - A Nature's Drugstore: An overview. *Int. Res. J. Biol. Sci*. **1**, 76-79.
149. Pathak, C.S. and Tiwari, S.K. 2010. Toxicological effects of neem *Azadirachta indica* A. Juss leaf powder against the ontogeny of *Corcyra cephalonica* (Staint.) (Lepidoptera: Pyralidae). *J. Biopest*. **3**, 617-621.
150. Faccin-Galhardi, L.C., Yamamoto, K.A., Ray, S., Ray, B., Linhares, R.E. and Nozawa, C. 2012. The *in vitro* antiviral property of *Azadirachta indica* polysaccharides for poliovirus. *J. Ethnopharmacol*. **142**, 86-90.
151. Badam, L., Joshi, S.P. and Bedekar, S.S. 1999. *In vitro* antiviral activity of neem (*Azadirachta indica*. A. Juss) leaf extract against group B Coxsackieviruses. *J. Commun Dis*. **31**, 79-90.
152. Bansod, M.S. and Harle, U.N. 2009. *Vitex negundo* L.: phytochemical constituents, traditional uses and pharmacological properties: comprehensive review. *Pharmacologyonline Newsletter*. **1**, 286-302.
153. Kothandan, S. and Swaminathan, R. 2014. Evaluation of *in vitro* antiviral activity of *Vitex Negundo* L., *Hyptis suaveolens* (L) poit., *Decalepis hamiltonii* Wight & Arn., to Chikungunya virus. *Asian Pac. J. Trop. Dis*. **4**, 111-115.
154. Kannan, M., Rajendran, P., Vedha, V., Ashok, G., Anushka, S., Chandran, R.N.P. 2012. HIV-1 reverse transcriptase inhibition by *Vitex negundo* L. leaf extract and quantification of flavonoids in relation to anti-HIV activity. *J Cell Mol Biol*. **10**, 53-59.
155. Muthukrishnan, S., Palanisamy, S., Subramanian, S., Selvaraj, S., Mari, K.R. and Kuppulingam, R. Phytochemical Profile of *Erythrina variegata* by Using High-Performance Liquid Chromatography and Gas Chromatography-Mass Spectroscopy Analyses. *J. Acupunct. Meridian Stud*. **9**, 207-212.
156. Kumar, A., Lingadurai, S., Jain, A. and Barman, N.R. 2010. *Erythrina variegata* Linn: A review on morphology, phytochemistry, and pharmacological aspects. *Pharmacog Rev*. **4**, 147-152.
157. Muthukrishnan, S., Palanisamy, S., Santhanam, I. and Kaveriyappan, G. 2014. Phytochemical screening and antimicrobial activity of *Erythrina variegata*. *World J. Pharm. Pharmaceut. Sci*. **3**, 680-690.
158. Choudhary, N. and Singh V. 2018. A census of *P. longum*'s phytochemicals and their network pharmacological evaluation for identifying novel drug-like molecules against various diseases, with a special focus on neurological disorders. *PLOS ONE*. **13**, 1-38.
159. Nahak, G. and Sahu, R.K. 2011. Phytochemical evaluation and antioxidant activity of *Piper cubeba* and *Piper nigrum*. *J. Appl. Pharm. Sci*. **1**, 153-157.
160. Jiang, Z.Y., Liu, W.F., Zhang, X.M., Luo, J., Ma, Y.B. and Chen, J.J. 2013. Anti-HBV active constituents from *Piper longum*. *Bioorg Med Chem Lett*. **23**, 2123-2127.
161. Mukherjee, P.K. 2019. Antiviral evaluation of herbal drugs. Quality control and evaluation of herbal drugs. *Quality Control and Evaluation of Herbal Drugs*. **2019**, 599-628.
162. Goyal, B.M., Bansal, P., Gupta, V., Kumar, S., Singh, R. and Maithani, M. 2010. Pharmacological Potential of *Boerhaavia diffusa*: An Overview. *Int. J. Pharma. Sci. Drug Res*. **2**, 17-22.
163. Anbazhagan, G.K., Palaniyandi, S. and Joseph, B. 2019. Antiviral plant extracts. *Plant Extracts*, Edt. by Aman Dekebo.
164. Awasthi, L.P. and Menzel, G. 1986. Effect of root extract from *Boerhaavia diffusa* L., containing an antiviral principle upon plaque formation of RNA bacteriophages. *Zentralbl Mikrobiol*. **141**, 415-419.
165. Thirunavoukkarasu, M. and Nayak, P. 2016. A review of the plant *Boerhaavia diffusa*: its chemistry, pharmacology and therapeutical potential. *The J. Phytopharmacol*. **5**, 83-92.
166. Helal, I., Galala, A., Saad, H.E. and Halim, A. *Bioactive Constituents from Apium leptophyllum* Fruits. *Br. J. Pharm. Res*. **14**, 1-8.
167. Halim, A.F., Helal, I.E., Galala, A.A. and Saad, H.E.A. 2015. Chemical Composition and α -Amylase Inhibitory Activity of *Apium Leptophyllum* essential oils. *J. American Sci*. **11**, 204-209.

168. Sahoo, H.B., Patro, S.K., Sagar, R. and Santani, D.D. 2015. Mutagenic evaluation and spectroscopic characterization of flavonoidal fraction of *Apium leptophyllum* (Pers.) fruit. *Int J Nutr. Pharmacol. Neurol. Dis.* **5**, 82-88.
169. Arya, D. and Patni, V. 2013. Pharmacognostic profile and phytochemical investigation of *Pluchea lanceolata* Oliver & Hiern. *In vivo and in vitro*. *Int. J. Pharm. Sci. Rev. Res.* **22**, 157-161.
170. Ameyaw, Y. and Duker-eshun, G. 2009. The Alkaloid Contents of the Ethno-Plant Organs of Three Antimalarial Medicinal Plant Species in the Eastern Region of Ghana. *Int. J. Chem. Sci.* **7**, 48-58.
171. Daniel, V.N., Daniang, I.E. and Nimyel, N.D. 2011. Phytochemical analysis and mineral elements composition of *Ocimum basilicum* obtained in jos metropolis, plateau state, Nigeria. *Int. J. Eng. Technol.* **11**, 161-165.
172. Srivastava, P. and Shanker, K. 2012. *Pluchea lanceolata* (Rasana): Chemical and biological potential of Rasayana herb used in traditional system of medicine. *Fitoterapia.* **83**, 1371-1385.
173. Ried, K. and Fakler, P. 2014. Potential of garlic (*Allium sativum*) in lowering high blood pressure: mechanisms of action and clinical relevance. *Integr. Blood Press Contr.* **7**, 71-82.
174. Shu, S.N., Fang, F. and Dong, Y.S. 2003. An experimental study on the effect of allitridin on inhibiting the expression of HCMV immediate-early antigens *in vitro*. *Zhongguo Zhongyao Zazhi.* **28**, 969-970.
175. Tsai, Y., Cole, L.L., Davis, L.E., Lockwood, S.J., Simmons, V. and Wild, G.C. 1985. Antiviral properties of garlic: in vitro effects on influenza B, herpes simplex and coxsackie viruses. *Planta Med.* **5**, 460-461.
176. Zhen, H., Fang, F., Ye, D.Y., Shu, S.N., Zhou, Y.F. and Dong, Y.S. 2006. Experimental study on the action of allitridin against human cytomegalovirus in vitro: Inhibitory effects on immediate-early genes. *Antiviral. Res.* **72**, 68-74.
177. Rani, N.Z.A., Husain, K. and Kumolosasi, E. 2018. Moringa genus: A review of phytochemistry and pharmacology. *Front Pharmacol.* **9**, 1-26.
178. Gopalakrishnan, L., Doriya, K. and Kumar, D.S. 2016. Moringa oleifera: A review on nutritive importance and its medicinal application. *Food Sci. Human wellness.* **5**, 49-56.
179. Biswas, D., Nandy, S., Mukherjee, A., Pandey, D.K. and Dey, A. 2020. *Moringa oleifera* Lam. and derived phytochemicals as promising antiviral agents: A review. *South African J. Bot.* **129**, 272-282.
180. Ali, G.H., El-Taweel, G.E. and Ali, M.A. 2004. The cytotoxicity and antimicrobial efficiency of *Moringa oleifera* seeds extracts. *Int. J. Environ. Stud.* **61**, 699-708.
181. Nasr-Eldin, M., Abdelhamid, A. and Baraka, D. 2018. Antibiofilm and Antiviral Potential of Leaf Extracts from *Moringa oleifera* and Rosemary (*Rosmarinus officinalis* Lam.). *Egypt J. Microbiol.* **52**, 129-139.
182. Nworu, C. S., Okoye, E. L. and Ezeifeke, G. O. 2013. Extracts of *Moringa oleifera* Lam. showing inhibitory activity against early steps in the infectivity of HIV-1 lentiviral particles in a viral vector-based screening. *Afr J Biotechnol.* **12**, 4866-4873.
183. Imran, I., Altaf, I., Ashraf, M., Javeed, A., Munir, N. and Bashir, R. 2016. *In vitro* evaluation of antiviral activity of leaf extracts of *Azadirachta indica*, *Moringa oleifera*, and *Morus alba* against the foot and mouth disease virus on BHK-21 cell line. *Science Asia.* **42**, 392-396.
184. Chollom, S.C., Agada, G.O., Gotep, J.G., Mwankon, S.E., Dus, P.C., Bot, Y.S., Nyango, D.Y., Singnap, C.L., Fyaktu, E.J. and Okwori, A.E. 2012. Investigation of aqueous extract of *Moringa oleifera* lam seed for antiviral activity against newcastle disease virus in vivo. *J. Med. Plants Res.* **6**, 3870-3875.
185. Jabbar, S., Khan, M.T.H., Choudhuri, M.S.K. and Sil, B.K. 2004. Bioactivity studies of the individual ingredients of the Dashamularishta. *Pak. J. Pharm. Sci.* **17**, 9-17.
186. Bhattacharjee, A., Shashidhara, S.C. and Saha, S. 2013. Phytochemical and ethno-pharmacological profile of *Desmodium gangeticum* (L.) DC.: A review. *Int. J. Biomed. Res.* **4**, 507-515.
187. Karthikeyan, K., Selvam, G.S., Srinivasan, R., Chandran, C. and Kulothungan, S. 2012. *In vitro* antibacterial activity of *Desmodium gangeticum* (L.) DG. *Asian Pac. J. Trop. Dis.* **2**, 421-424.
188. Gayathri, P. and Kanakamany, M.T. 2018. Influence of storage environment and packing materials on seed germination and viability of *Desmodium gangeticum* (L.) DC. *J. Med. Aroma. Plants.* **9**, 23-33.
189. Al-Snafi, A.E. 2015. The pharmacological importance of *Brassica nigra* and *Brassica rapa* grown in Iraq. *J. Pharmaceutical Biol.* **5**, 240-253.
190. Upwar, N.K., Patel, R., Waseem, N. and Mahobia, N.K. 2011. In vitro anthelmintic activity of *Brassica nigra* Linn. seeds. *Int. J. Nat. Prod. Res.* **1**, 1-3.
191. Ali, A.M.H. and Mohamed, A.E. 2018. Antifeedant effect of *Brassica nigra* seeds against cotton leafworm Spodoptera littoralis and its potential antibacterial activity. *GSC Biol. Pharmaceut. Sci.* **5**, 32-40.
192. Danlami, U. 2016. Phytochemical, Nutritional and Antimicrobial Evaluations of the Aqueous Extract of *Brassica Nigra* (Brassicaceae) Seeds. *American J. App. Chem.* **4**, 161-163.
193. Jain, V. and Verma, S.K. 2012. Pharmacology of *Bombax ceiba* Linn. Vol. 12. Published by SpringerBriefs.
194. Kumar, A. and Shamim, S. 2018. Evaluation of wound healing activity of leaves of *Bombax ceiba*. *J. Med. Sci. Clin. Res.* **6**, 119-126.
195. Shah, S.S., Shah, S.S., Iqbal, A., Ahmed, S., Khan, W.M., Hussain, S. and Li, Z. Phytochemical screening and antimicrobial activities of red silk cotton tree (*Bombax ceiba* L.). *Pakistan J. Pharmaceut. Sci.* **31**, 947-952.

196. Siddiqua, A., Zahra, M., Begum, K. and Jamil, M. 2018. The Traditional Uses, Phytochemistry and Pharmacological Properties of *Cassia fistula*. *J. Pharm. Pharmacol. Res.* **2**, 15-23.
197. Bhalodia, N.R. and Shukla, V.J. 2011. Antibacterial and antifungal activities from leaf extracts of *Cassia fistula* L.: An ethnomedicinal plant. *J. Adv. Pharm. Technol. Res.* **2**, 104-109.
198. Rahman, M.A., Sultana, P., Islam, M.S., Mahmud, M.T., Rashid, M.M.O. and Hossen, F. 2016. Comparative Antimicrobial Activity of *Areca catechu* Nut Extracts using Different Extracting Solvents. *Bangladesh J. Microbiol.* **31**, 19-23.
199. Patil, P.R., Rakesh, S.U., Dhabale, P.P.N. and Burade, P.K.B. 2009. Pharmacological activities of *Areca catechu* Linn. A Review. *J. Pharm. Res.* **2**, 683-687.
200. Xiao, Y., Yang, Y., Yong, J. and Lu, C. 2019. Chemical Components and Biological Activities of *Areca catechu* L. *Biomed. Res. Rev.* **8**, 8-12.
201. Peng, W., Liu, Y.J., Wu, N., Sun, T., He, X.Y. and Gao, Y.X. 2015. *Areca catechu* L. (Arecaceae): A review of its traditional uses, botany, phytochemistry, pharmacology and toxicology. *J. Ethnopharmacol.* **164**, 340-356.
202. Hassan, W., Kazmi, S.Z., Noreen, H., Riaz, A. and Zaman, B. 2016. Antimicrobial activity of *cinnamomum tamala* leaves. *J Nutr. Disord. Ther.* **6**, 1-5.
203. Howlader, M.S.I., Dey, S.K., Hira, A. and Ahmed, A. 2012. Evaluation of antinociceptive, antimicrobial and cytotoxic activities of ethanolic extract of *cinnamomum tamala* leaves from Bangladesh. *Pharmacologyonline.* **2**, 73-81.
204. Kumar, S., Sharma, S. and Vasudeva, N. 2012. Chemical compositions of *Cinnamomum tamala* oil from two different regions of India. *Asian Pac J Trop Dis.* **2**, 761-764.
205. Kumar, A., Rahal, A., Chakraborty, S., Tiwari, R., Latheef, S.K. and Dhama, K. 2013. *Ocimum sanctum* (Tulsi): a miracle herb and boon to medical science – A Review. *Int J Agr. Plant Prod.* **4**, 1580-1589.
206. Borah, R. and Biswas, S.P. 2018. Tulsi (*Ocimum sanctum*), excellent source of phytochemicals. *Int. J. Environ. Agri. Biotech.* **3**, 1732-1738.
207. Jayati, B.A.K., Kumar, A., Goel, A., Gupta, S. and Rahal, A. 2013. *In vitro* antiviral potential of *Ocimum sanctum* leaves extract against Newcastle disease virus of poultry. *Int. J. Microbiol. Immunol. Res.* **2**, 51-55.
208. Ghoke, S.S., Sood, R., Kumar, N., Pateriya, A.K., Bhatia, S. and Mishra, A. 2018. Evaluation of antiviral activity of *Ocimum sanctum* and *Acacia arabica* leaves extracts against H9N2 virus using embryonated chicken egg model. *BMC Complement Altern. Med.* **18**, 174-184.
209. Saleem, T.M., Chetty, C., Ramkanth, S., Alagusundaram, M., Gnanaprakash, K., Rajan, V.T. and Angalapameswari, S. 2009. *Solanum nigrum* Linn.-A review. *Pharmacog. Rev.* **3**, 342-345.
210. Javed, T., Ashfaq, U.A., Riaz, S., Rehman, S. and Riazuddin, S. 2011. In-vitro antiviral activity of *Solanum nigrum* against Hepatitis C Virus. *Virol. J.* **8**, 26-34.
211. Chauhan, A., Saini, R., Shori, A. and Dwivedi, J. 2012. *Solanum nigrum* with dynamic therapeutic role: A review. *Int J. Pharm. Sci. Rev. Res.* **15**, 65-71.
212. Lampariello, L., Cortelazzo, A., Guerranti, R., Sticozzi, C. and Valacchi, G. 2012. The Magic Velvet Bean of *Mucuna pruriens*. *J Trad Complement Med.* **2**, 331-339.
213. Salau, A.O. and Odeleye, O.M. 2007. Antimicrobial activity of *Mucuna pruriens* on selected bacteria. *Afr J Biotechnol.* **6**, 2091-2092.
214. Srivastava, A.K., Naseer, A. and Gupta, A. 2022. Comparative Assessment of Polyherbal Formulation and *Mucuna pruriens* Extract as Neuroprotectant by Using MPTP Screening Mouse Model of Parkinson's Disease. *Curr. Bioact. Comp.* **18**, 78-86.
215. Borlinghaus, J., Albrecht, F., Gruhlke, M.C.H., Nwachukwu, I.D. and Slusarenko, A.J. 2014. Allicin: Chemistry and biological properties. *Molecules.* **19**, 12591-12618.
216. Mahalik, G., Jali, P., Sahoo, S. and Satapathy, K.B. 2020. Ethnomedicinal, phytochemical and pharmacological properties of *Mangifera indica* L: A review. *Int. J. Bot. Stud.* **5**, 1-5.
217. Xiang, Y.F., Ju, H.Q., Li, S., Zhang, Y.J., Yang, C.R. and Wang, Y.F. 2010. Effects of 1,2,4,6-tetra-O-galloyl- β -D-glucose from *P. emblica* on HBsAg and HBeAg secretion in HepG2.2.15 cell culture. *Virol. Sin.* **25**, 375-380.
218. Mirjalili, M.H., Fakhr-Tabatabaei, S.M., Alizadeh, H., Ghassempour, A. and Mirzajani, F. 2009. Genetic and withaferin A analysis of Iranian natural populations of *Withania somnifera* and *W. coagulans* by RAPD and HPTLC. *Nat. Prod. Commun.* **4**, 337-346.
219. Guan, S., Xu, Y., Qiao, Y., Kuai, Z., Qian, M. and Jiang, X. 2018. A novel small molecule displays two different binding modes during inhibiting H1N1 influenza A virus neuraminidases. *J. Struct. Biol.* **202**, 142-149.
220. Nelson, K.M., Dahlin, J.L., Bisson, J., Graham, J., Pauli, G.F. and Walters, M.A. 2017. The Essential Medicinal Chemistry of Curcumin. *J. Med. Chem.* **60**, 1620-1637.
221. Matei, E., Basu, R., Furey, W., Shi, J., Calnan, C. and Aiken, C. 2016. Structure and glycan binding of a new cyanovirin-N homolog. *J. Biol. Chem.* **291**, 18967-18976.
222. Miękus, N., Marszałek, K., Podlacha, M., Iqbal, A., Puchalski, C. and Świergiel, A.H. 2020. Health benefits of plant-derived sulfur compounds, glucosinolates and organosulfur compounds. *Molecules* **25**, 3804-3826.